U.S. DEPARTMENT OF COMMERCE PATENT AND TRADEMARK OFFICE FORM-PTO-1390 ATTORNEY'S DOCKET NUMBER (Rev. 12-29-99) TRANSMITTAL LETTER TO THE UNITED STATES 012627-025 DESIGNATED/ELECTED OFFICE (DO/EO/US) U.S. APPLICATION NO. (If known, see 37 C.F.R. 1.5) CONCERNING A FILING UNDER 35 U.S.C. 371 INTERNATIONAL APPLICATION NO. INTERNATIONAL FILING DATE PRIORITY DATE CLAIMED PCT/EP00/02330 21 SEPTEMBER 2000 19 MARCH 1999 31 DECEMBER 1999 TITLE OF INVENTION Method for Identifying Organisms by Means of Comparative Genetic Analysis and Primers and Hybridization Probes for Carrying out this Method APPLICANT(S) FOR DO/EO/US Hans SCHACKERT et al. Applicant herewith submits to the United States Designated/Elected Office (DO/EO/US) the following items and other information: This is a FIRST submission of items concerning a filing under 35 U.S.C. 371. 2. This is a SECOND or SUBSEQUENT submission of items concerning a filing under 35 U.S.C. 371. This is an express request to begin national examination procedures (35 U.S.C. 371(f)) at any time rather than delay examination 3. until the expiration of the applicable time limit set in 35 U.S.C. 371(b) and the PCT Articles 22 and 39(1). A proper Demand for International Preliminary Examination was made by the 19th month from the earliest claimed priority date. A copy of the International Application as filed (35 U.S.C. 371(c)(2)) is transmitted herewith (required only if not transmitted by the International Bureau). has been transmitted by the International Bureau. is not required, as the application was filed in the United States Receiving Office (RO/US) A translation of the International Application into English (35 U.S.C. 371(c)(2)). Amendments to the claims of the International Application under PCT Article 19 (35 U.S.C. 371(c)(3)) 7. are transmitted herewith (required only if not transmitted by the International Bureau). have been transmitted by the International Bureau. have not been made; however, the time limit for making such amendments has NOT expired.  $\bowtie$ have not been made and will not be made. A translation of the amendments to the claims under PCT Article 19 (35 U.S.C. 371(c)(3)). An oath or declaration of the inventor(s) (35 U.S.C. 371(c)(4)). A translation of the annexes to the International Preliminary Examination Report under PCT Article 36 (35 U.S.C. 371(c)(5)). Items 11. to 16. below concern other document(s) or information included: An Information Disclosure Statement under 37 CFR 1.97 and 1.98. An assignment document for recording. A separate cover sheet in compliance with 37 CFR 3.28 and 3.31 is included. 12. A FIRST preliminary amendment. 13. A SECOND or SUBSEQUENT preliminary amendment. A substitute specification. A change of power of attorney and/or address letter. Other items or information: International Search Report International Preliminary Examination Report PCT Request form (PCT/RO/101) (8) sheets of Drawings (48) Sheets (List of species sequences)

INTERNATIONAL APPLICATION NO. ATTORNEY'S DOCKET NUMBER U.S. APPLICATION NO (If known 936738 012627-025 PCT/EP00/02330 **UNASSIGNED** PTO USE ONLY **CALCULATIONS** The following fees are submitted: Basic National Fee (37 CFR 1.492(a)(1)-(5)): Neither international preliminary examination fee (37 CFR 1.482) nor international search fee (37 CFR 1.445(a)(2)) paid to USPTO and International Search Report not prepared by the EPO or JPO ...... \$1,000.00 (960) International preliminary examination fee (37 CFR 1.482) not paid to USPTO but International Search Report prepared by the EPO or JPO . . . . . . . . \$860.00 (970) International preliminary examination fee (37 CFR 1.482) not paid to USPTO but international search fee (37 CFR 1.445(a)(2)) paid to USPTO . . . . . . . . . \$710.00 (958) International preliminary examination fee paid to USPTO (37 CFR 1.482) but all claims did not satisfy provisions of PCT Article 33(1)-(4) ..... \$690.00 (956) International preliminary examination fee paid to USPTO (37 CFR 1.482) and all claims satisfied provisions of PCT Article 33(1)-(4) . . . . . . . . . . . . . \$100.00 (962) ENTER APPROPRIATE BASIC FEE AMOUNT = 860.00 \$ 20 🗆 30 🗖 Surcharge of \$130.00 (154) for furnishing the oath or declaration later than months from the earliest claimed priority date (37 CFR 1.492(e)). Rate Number Filed Number Extra Claims X\$18.00 (966) 576.00 52 -20 = 32 **Total Claims** Ś 1440.00 X\$80.00 (964) 21 -3 = 18 Independent Claims + \$270.00 (968) ŝ Multiple dependent claim(s) (if applicable) 2016.00 TOTAL OF ABOVE CALCULATIONS = Ś Ś Reduction for 1/2 for filing by small entity, if applicable (see below). \$ 2876.00 SUBTOTAL = 20 🗆 30 🗀 ŝ Processing fee of \$130.00 (156) for furnishing the English translation later than months from the earliest claimed priority date (37 CFR 1.492(f)). TOTAL NATIONAL FEE = \$ 2876.00 Fee for recording the enclosed assignment (37 CFR 1.21(h)). The assignment must be accompanied by an appropriate cover sheet (37 CFR 3.28, 3.31). \$40.00 (581) per property + \$ 2876.00 TOTAL FEES ENCLOSED = Amount to be: refunded charged Small entity status is hereby claimed. а  $\boxtimes$ A check in the amount of \$ 2876.00 to cover the above fees is enclosed. b. Please charge my Deposit Account No. 02-4800 in the amount of \$\_\_\_\_\_ to cover the above fees. A duplicate copy of this sheet C. is enclosed. The Commissioner is hereby authorized to charge any additional fees which may be required, or credit any overpayment to Deposit  $\boxtimes$ d. Account No. 02-4800. A duplicate copy of this sheet is enclosed. NOTE: Where an appropriate time limit under 37 CFR 1.494 or 1.495 has not been met Appetition to revive (37 CFR 1.137(a) or (b)) must be filed and granted to restore the application to pending status. SEND ALL CORRESPONDENCE TO: Teresa Stanek Rae BURNS, DOANE, SWECKER & MATHIS, L.L.P. P.O. Box 1404 Alexandria, Virginia 22313-1404 Teresa Stanek Rea (703) 836-6620 NAME 30.427 REGISTRATION NUMBER

# JC16 Rec'd PCT/PTO SEP 1 7 2001

Patent Attorney's Docket No. <u>012627-025</u>

# IN THE UNITED STATES PATENT AND TRADEMARK OFFICE

In re Patent Application of	)
Prof. Dr. Hans SCHACKERT et al.	) Group Art Unit: Unassigned
Application No.: Unassigned (Corresponding to PCT/EP00/02330 International Filing Date: 16 March 2000	) Examiner: Unassigned ) ) )
For: METHOD FOR IDENTIFYING ORGANISMS BY MEANS OF COMPARATIVE GENETIC ANALYSIS AND PRIMERS AND HYBRIDISATION PROBES FOR CARRYING OUT THIS METHOD	) ) ) ) ) ) ) )

# PRELIMINARY AMENDMENT

Assistant Commissioner for Patents Washington, D.C. 20231

Sir:

Prior to examination, kindly amend the above-identified application as follows:

#### IN THE CLAIMS:

Kindly replace claims 1-45 and 47-52 as follows:

1. (Amended) A method of identifying organisms by comparative genetic analysis, wherein the coding and/or non-coding areas and/or functionally significant areas of highly conserved genes and/or their homologous genes and/or their cDNA copies and/or their pseudogenes are amplified using PCR and are subsequently genotyped and analyzed.

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- 2. (Amended) The method according to claim 1, wherein one primer pair each is used for each specific segment of the highly conserved gene, which is located in the highly conserved exon region and/or non-coding areas and/or functionally significant areas and/or in the 5'- or 3'-untranslated area of the gene and binds in as many studied species DNAs as possible, preferable in all studied species DNAs, and enables the amplification of the corresponding gene area.
- 3. (Amended) The method according to claim 1, wherein the coding and/or non-coding areas located between the primers and being either highly variant intron regions and/or variant exon regions or 5'- or 3'-untranslated areas of the gene, are analyzed as regards their sequence and identified by comparison with the species-specific sequence variants.
- 4. (Amended) The method according to claim 1, wherein either the sense strand or the antisense strand of any species DNA or also their PCR copies are used for the identification.
- (Amended) The method according to claim 1, wherein animals are identified.

- 6. (Amended) The method according to claim 1, wherein vertebrates are identified.
- 7. (Amended) The method according to claim 1, wherein mammals are identified.
  - 8. (Amended) The method according to claim 1, wherein plants are identified.
- 9. (Amended) The method according to claim 1, wherein genotying is carried out by DNA sequencing, any hybridization method, restriction fragment length analyses, chromatographic methods, spectroscopic and mass-spectroscopic methods, allele-specific PCR or by other methods suitable for detecting DNA sequence variants.
- 10. (Amended) The method according to claim 1, wherein exon and/or intron areas as well as functionally significant areas of the highly conserved tumor suppressor gene PTEN/MMAC1 and its homologues are used for amplification and subsequent genetic analysis.
- 11. (Amended) The method according to claim 1, wherein cDNA copies of the PTEN/MMAC1 gene and its homologues are used for the genetic analysis.

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- 12. (Amended) The method according to claim 1, wherein pseudogenes or segments of pseudogenes of the PTEN/MMAC1 gene and its homologues are used for the genetic analysis.
- 13. (Amended) The method according to claim 1, wherein exons arranged next to the PTEN/MMAC1 gene and its homologues and/or the parts of the introns following the exons are analyzed genetically.
- 14. (Amended) The method according to claim 1, wherein the exon regions 1 and 2 and/or 3 and 4 and/or 4 and 5 and/or 5 and 6 and/or 6 and 7 and/or 7 and 8 and/or 8 and 9 with the enclosed intron regions 1 and/or 2 and/or 3 and/or 4 and/or 5 and/or 6 and/or 7 and/or 8 as well as the 5'- and 3'-untranslated regions of the PTEN/MMAC1 gene and their homologues are used for the genetic analysis.
- 15. (Amended) The method according to claim 1, comprising selecting areas of highly conserved genes and/or pseudogenes and their homologues, constructing suitable oligonucleotides as primers which bind to the corresponding complementary coding and/or non-coding areas and/or functionally significant areas, amplifying them by means of a suitable technique and comparatively analyzing the sequence of the corresponding coding and/or non-coding area of various species by genetic analysis.

- 16. (Amended) The method according to claim 15, wherein areas of the PTEN/MMAC1 gene and/or the pseudogene and their homologues are selected.
- 17. (Amended) The method according to claim 15, wherein differing sequence segments of each individual exon, intron or untranslated region of the PTEN/MMAC1 gene and their homologues or the corresponding cDNA are selected.
- 18. (Amended) The method according to claim 1, wherein genotyping of pig DNA which is obtained from foodstuffs, is carried out on the basis of the gene sequence variant of PTEN/MMAC1 containing a 9-base pair long deletion.
- 19. (Amended) An oligonucleotide primer for the PCR and the sequencing of exon 1 and/or 5'-untranslated region of the PTEN/MMAC1 gene and its homologues, comprising the following sequences:

PTENex1-401 sense

5'-cccttctactgcctcca -3'

PTENex1 -465 sense

5'- gggagggggtctgagt -3'

PTENex1 ATG sense

5'- atgacagccatcatcaaaga -3'

PTENex1 R antisense

5'- aggtcaagtctaagtcgaatc -3'

20. (Amended) The oligonucleotide primer for PCR and the sequencing of exon 2 of the PTEN/MMAC1 gene and its homologues, comprising the following sequences:

PTENex2F sense

5'- atatttatccaaacattattgctat -3'

PTENex2R antisense

5'- cttactacatcatcatattgttcc -3'

21. (Amended) The oligonucleotide primer for PCR and the sequencing of exon 4, intron 4 and exon 5 of the PTEN/MMAC1 gene and its homologues, comprising the following sequences:

Zoo43sUV sense

5'- tgtgctgagagacattatgac -3'

SPL5 sense
5'- aaatttaattgcagaggt -3'

5'- ttgtctctggtccttacttc -3'

Zoo44aRV antisense

22. (Amended) The oligonucleotide primer for PCR and the sequencing of exon 5 of the PTEN/MMAC1 gene and its homologues, comprising the following sequences:

PTEN se sense

5'- atcttgaccaatggctaagtg -3'

Zoo44aRV antisense

5'- ttgtctctggtccttacttc -3'

23. (Amended) The oligonucleotide primer for PCR and the sequencing of exon 6 of the PTEN/MMAC1 gene and its homologues, comprising the following sequences:

PTENex6F sense

5'- gga gta act att ccc agt cag ag -3'

PTENex6R antisense

5'- gca agt tcc gcc act gaa -3'

24. (Amended) The oligonucleotide primer for PCR and the sequencing of exon 7 of the PTEN/MMAC1 gene and its homologues, comprising the following sequences:

PTENex7F sense

5'- cct cag ttt gtg gtc tgc ca -3'

PTENex7R antisense

5'- c ctt ttt tag cat ctt gtt ctg ttt -3'

25. (Amended) The oligonucleotide primer for PCR and the sequencing of exon 8 of the PTEN/MMAC1 gene and its homologues, comprising the following sequences:

PTENex8F sense

5'- caa aat gtt tca ctt ttg ggt aaa -3'

PTENex8R antisense

5'- taa aat ttg gag aaa agt atc ggt t -3'

26. (Amended) The oligonucleotide primer for PCR and the sequencing of exon 9 of the PTEN/MMAC1 gene and its homologues, comprising the following sequences:

PTENex9F sense

5'- gtg aag ctg tac ttc aca aaa ac -3'

PTENex9tga antisense

5'- aaa aaa att cag act ttt gta att tg -3'

- 27. (Amended) The method according to claim 1, wherein the DNA amplification involves a mixture of oligonucleotides which differ at the 3' region of the oligonucleotide as regards its length by one or more nucleotides or which differ as regards their nucleotide sequence at the 3' end of the oligonucleotide at one or more positions.
  - 28. (Amended) The method according to claim 1, wherein the oligonucleotides

sense:

5'- cga cgt tgt aaa acg acg gcc agt tgt gct gag aga cat tat gac -3',

5'- cga cgt tgt aaa acg acg gcc agt tgt gct gag aga cat tat -3',

5'- cga cgt tgt aaa acg acg gcc agt tgt gct gag aga cat t -3',

antisense:

5' - cag gaa aca get atg act tgt etc tgg tec tta ett e -3',

5'- cag gaa aca get atg act tgt etc tgg tee tta e -3',

5'- cag gaa aca get atg act tgt ete tgg tee t -3'

are used for the amplification.

- 29. (Amended) The method according to claim 1, wherein the oligonucleotides sense:
- 5'- cga cgt tgt aaa acg acg gcc agt tgt gct gag aga cat tat gaa -3',
- 5'- cga cgt tgt aaa acg acg gcc agt tgt gct gag aga cat tat gac -3',
- 5'- cga cgt tgt aaa acg acg gcc agt tgt gct gag aga cat tat gag -3',
- 5'- cga cgt tgt aaa acg acg gcc agt tgt gct gag aga cat tat gat -3',

antisense:

- 5'- cag gaa aca get atg act tgt etc tgg tee tta ett a -3',
- 5'- cag gaa aca get atg act tgt etc tgg tee tta ett e -3',
- 5'- cag gaa aca get atg act tgt etc tgg tec tta ett g -3',
- 5'- cag gaa aca get atg act tgt etc tgg tee tta ett t -3'

are used for the amplification.

30. (Amended) The method according to claim 1, wherein DNA sequencing methods are used for the genetic analysis.

- 31. (Amended) The method according to claim 1, wherein DNA sequencing techniques are used in the genetic analysis for the PTEN/MMAC1 and/or its pseudogenes and their homologues.
- 32. (Amended) The method of distinguishing the DNA of various species, wherein at least one hybridization probe pair is used, the melting points of different combinations are determined and compiled for each species into a panel.
- 33. (Amended) The method of distinguishing the DNA of various species, wherein at least one hybridization probe pair is used and at least one gene segment is amplified, differing hybridization probe pairs hybridize to different gene segments, and the melting points of the different combinations are determined and compiled for each species into a panel and/or compared with this panel for the purpose of identification.
- 34. (Amended) The method of distinguishing the DNA of different species according to claim 33, wherein at least one hybridization probe pair is used and at least one gene segment of at least one species is amplified, differing hybridization probe pairs hybridize to different gene segments of various species, and the melting points of the different combinations are determined and compiled for each species into a panel and/or compared with this panel for the purpose of identification.

- 35. (Amended) The method of distinguishing the DNA of various species according to claim 33, wherein at least two hybridization probes of SEQ Nos. 3 to 8 are used, the melting points of different combinations are determined and compiled for each species into a panel.
- 36. (Amended) The method according to claim 33, wherein the species differentiation of pig DNA from various other species is made using the hybridization probe pair Al/A2 as the hybridization probe pair.
- 37. (Amended) The method according to claim 33, wherein the hybridization probes are used in combinations Cl/C2; A1/B2; A1/A2; C1/A2; B1/B2; B1/A2 for the species differentiation between various species.
- 38. (Amended) LightCycler hybridization probes for exon 5, comprising the sequences:

A1: 5'- tgc ata ttt gtt tca tcc ggg caa att -fluorescein -3'

A2: 5'- LC Red 705 - tta aag gca caa gat ttc tat ggg ga - ph -3'

B1: 5'- tgc ata ttt att aca tcg ggg caa att -fluorescein -3'

B2: 5'- LC Red 640 - aag gca caa gag gcc cta gat ttc ta - ph -3'

C1: 5'-tgc ata ttt gtt aca tcg ggg taa att fluorescein -3'

C2: 5'- LC Red 640 - aag gca caa gag gcc cta gat ttc ta - ph -3'

39. (Amended) LightCycler hybridization probes for exon 6, comprising the sequences

PTENex6FL

5'- tca tct gga tta tag acc agt ggc act - fluorescein -3'

PTENex6LC 640

5'- LC Red 640 - ttc aca aga tga tgt ttg aaa cta ttc caa- ph -3'

PTENex6F\*

5'- gtg cca ctg gtc tat aat cca gat- fluorescein -3'

PTENex6L\* 705

5'- LC Red 705- ttc ttt aac agg tag cta taa taa tac aca ta- ph -3'

40. (Amended) LightCycler hybridization probes for exon 7, comprising the sequences

PTENex7F\*

5'- taa agg tga aga tat att cct cca att ca - fluorescein -3'

PTENex7L\*640

5'-LC Red 640- acc cac acg acg gga aga caa g - ph -3'

PTENex7 FL

5'-ggtaacggctgagggaactcaaagtac - fluorescein -3'

PTENex7 LC (705-labeled)

5'-LC Red 705- tgaacttgtcttcccgtcgtgtgg- ph -3'

41. (Amended) LightCycler hybridization probes for exon 8, comprising the following sequences

PTENex8F\*

5'- tga caa gga ata tct agt act tac ttt aac aaa-fluorescein -3'

PPTENex8L\* 705

5'-LC Red 705 - ctt gac aaa gca aat aaa gac aaa gc- ph -3'

PTENex8 FLU

5' - tgctatcgatttcttgatcacatagacttccatttt - fluorescein -3'

PTENex8 LCR (640-labeled)

5'-LC Red 640- actttttctgaggtttcctctggtcctggtat - ph -3'

42. (Amended) [The] LightCycler hybridization probes for exon 9, comprising the following sequences

PTENex9 FL

5'-aac atc tgg tgt tac aga agt tga act gct- fluorescein -3'

PTENex9 LC 640

5'-LC-640- cct ctg gat ttg acg gct cct cta ct - ph -3'

43. (Amended) Hybridization probe pair AI/A2: specific to PTEN pseudogene pig, comprising

SEQ No. 3 A1: 5'- tgc ata ttt gtt tca tcc ggg caa att -fluorescein -3'

SEQ No. 4 A2: 5'-LC Red 705- tta aag gca caa gat ttc tat ggg ga - ph -3'

44. (Amended) Hybridization probe pair B1/B2: specific to pseudogene man, comprising

SEQ No. 5 B1: 5'- tgc ata ttt att aca tcg ggg caa att -fluorescein -3'

SEQ No. 6 B2: 5'-LC Red 640- aag gca caa gag gcc cta gat ttc ta -ph -3'

45. (Amended) Hybridization probe pair C1/C2: specific to PTEN pseudogene man (C2) and homologue of pig (C1) comprising

SEQ No. 7 - C1: 5'- tgc ata ttt gtt aca tcg ggg taa att - fluorescein -3'

SEQ No. 8 - C2: corresponds to probe B2.

- 47. (Amended) DNA sequences of homologues of the PTEN/MMAC1 gene and/or of homologues of the PTEN/MMAC1 pseudogene, which are compiled in the annex under "list of species sequences", which as compared to the PTEN/MMAC1 gene and/or the PTEN/MMAC1 pseudogene comprise genetic variants comprising base substitutions and/or insertions and/or deletions and are suited for identifying corresponding species.
- 48. (Amended) A kit for carrying out the method according to claim 1, comprising:
  - a) one or more vessels comprising PCR and/or sequencing oligonucleotides binding to highly conserved genes, the oligonucleotides being optionally labeled radioactively or by means of a dye or in another way,
  - b) vessels having further common reagents for DNA amplification and/or DNA analysis,

and

- a vessel containing a control DNA which is suited for testing the oligonucleotides and the reaction conditions.
- 49. (Amended) The kit according to claim 48, comprising:
- a) one or more vessels with PCR and/or sequencing oligonucleotides.

- 50. (Amended) The kit for identifying species for carrying out the method according to claim 1, comprising:
  - a) a vessel having an oligonucleotide pair comprising the following sequences:

    5'- cga cgt tgt aaa acg acg gcc agt tgt gct gag aga cat tat gac -3' and 5'- cag
    gaa aca gct atg act tgt ctc tgg tcc tta ctt c -3',
  - b) two vessels with one of the following sequencing oligonucleotides each,
    these oligonucleotides being optionally labeled radioactively or by means of
    a dye or in another way:
    - 5'- cag gaa aca gct atg ac -3' and
    - 5'- cga cgt tgt aaa acg acg gcc agt -3',
  - c) a vessel containing a control *DNA*, which is suited for testing the oligonucleotides and the reaction conditions.
- 51. (Amended) The kit (Light Cycler Kit) for carrying out the method according to claim 32, comprising
  - a) one or more vessels containing PCR primers and hybridization probes, which bind to highly conserved genes, the hybridization probes being optionally labeled by means of a dye,
  - b) vessels containing further common reagents for DNA amplification and/or DNA analysis,

and

- a vessel containing a control DNA which is suited for testing the oligonucleotides and the reaction conditions.
- 52. (Amended) The kit (Light Cycler Kit) for carrying out the method according to claim 32, comprising:
  - a) one or more vessels with PCR primers and hybridization probes.

## **REMARKS**

Entry of the foregoing amendments is respectfully requested.

Respectfully submitted,

BURNS, DOANE, SWECKER & MATHIS, L.L.P.

By:

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Date: September 17, 2001

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# **Attachment to Preliminary Amendment dated XXX**

#### Marked-up Claims 1-45 and 47-52

- 1. (Amended) A method of identifying organisms by comparative genetic analysis, [characterized in that] wherein the coding and/or non-coding areas and/or functionally significant areas of highly conserved genes and/or their homologous genes and/or their cDNA copies and/or their pseudogenes are amplified using PCR and are subsequently genotyped and analyzed.
- 2. (Amended) The method according to claim 1, [characterized in that] wherein one primer pair each is used for each specific segment of the highly conserved gene, which is located in the highly conserved exon region and/or non-coding areas and/or functionally significant areas and/or in the 5'- or 3'-untranslated area of the gene and binds in as many studied species DNAs as possible, preferable in all studied species DNAs, and enables the amplification of the corresponding gene area.
- 3. (Amended) The method according to [claims 1 and 2] claim 1, [characterized in that] wherein the coding and/or non-coding areas located between the primers and being either highly variant intron regions and/or variant exon regions or 5'- or 3'-untranslated areas of the gene, are analyzed as regards their sequence and identified by comparison with the species-specific sequence variants.

#### **Attachment to Preliminary Amendment dated XXX**

#### Marked-up Claims 1-45 and 47-52

- 4. (Amended) The method according to [claims 1 to 3] <u>claim 1</u>, [characterized in that] <u>wherein</u> either the sense strand or the antisense strand of any species DNA or also their PCR copies are used for the identification.
- 5. (Amended) The method according to [claims 1 to 4] <u>claim 1</u>, [characterized in that preferably] <u>wherein</u> animals are identified.
- 6. (Amended) The method according to [claims 1 to 4] <u>claim 1</u>, [characterized in that preferably] <u>wherein</u> vertebrates are identified.
- 7. (Amended) The method according to [claims 1 to 4] <u>claim 1</u>, [characterized in that preferably] <u>wherein mammals are identified</u>.
- 8. (Amended) The method according to [claims 1 to 4] <u>claim 1</u>, [characterized in that preferably] <u>wherein plants are identified</u>.
- 9. (Amended) The method according to [claims 1 to 4] <u>claim 1</u>, [characterized in that] <u>wherein</u> genotying is carried out by DNA sequencing, any hybridization [methods] method, restriction fragment length analyses, chromatographic methods, spectroscopic and

#### Attachment to Preliminary Amendment dated XXX

## Marked-up Claims 1-45 and 47-52

[in particular] mass-spectroscopic methods, allele-specific PCR or by other methods suitable for detecting DNA sequence variants.

- 10. (Amended) The method according to [claims 1 to 4] <u>claim 1</u>, [characterized in that] <u>wherein</u> exon and/or intron areas as well as functionally significant areas of the highly conserved tumor suppressor gene PTEN/MMAC1 and its homologues are used for amplification and subsequent genetic analysis.
- 11. (Amended) The method according to [claims 1 to 4] <u>claim 1</u>, wherein cDNA copies of the PTEN/MMAC1 gene and its homologues are used for the genetic analysis.
- 12. (Amended) The method according to [claims 1 to 4] <u>claim 1</u>, wherein pseudogenes or segments of pseudogenes of the PTEN/MMAC1 gene and its homologues are used for the genetic analysis.
- 13. (Amended) The method according to [claims 1 to 4] <u>claim 1</u>, [characterized in that preferably] <u>wherein exons arranged [side by side of] next to the PTEN/MMAC1 gene and its homologues and/or the parts of the introns following the exons are analyzed genetically.</u>

# Attachment to Preliminary Amendment dated XXX

#### Marked-up Claims 1-45 and 47-52

- 14. (Amended) The method according to [claims 1 to 4] claim 1, [characterized in that] wherein the exon regions 1 and 2 and/or 3 and 4 and/or 4 and 5 and/or 5 and 6 and/or 6 and 7 and/or 7 and 8 and/or 8 and 9 with the enclosed intron regions 1 and/or 2 and/or 3 and/or 4 and/or 5 and/or 6 and/or 7 and/or 8 as well as the 5'- and 3'-untranslated regions of the PTEN/MMAC1 gene and their homologues are used for the genetic analysis.
- by] comprising selecting areas of highly conserved genes and/or pseudogenes and their homologues, constructing suitable oligonucleotides as primers which bind to the corresponding complementary coding and/or non-coding areas and/or functionally significant areas, amplifying them by means of a suitable technique and comparatively analyzing the sequence of the corresponding coding and/or non-coding area of various species by genetic analysis.
- 16. (Amended) The method according to claim 15, [characterized in that] wherein areas of the PTEN/MMAC1 gene and/or the pseudogene and their homologues are selected.
- 17. (Amended) The method according to [claims 15 and 16] claim 15, [characterized in that] wherein differing sequence segments of each individual exon, intron R:\012627\025\Docs\01-09-12 amend.wpd

#### Attachment to Preliminary Amendment dated XXX

# Marked-up Claims 1-45 and 47-52

or untranslated region of the PTEN/MMAC1 gene and their homologues or the corresponding cDNA are selected.

- 18. (Amended) The method according to [claims 1 to 17] claim 1, [characterized in that] wherein genotyping of pig DNA which is obtained [preferably] from foodstuffs, is carried out on the basis of the gene sequence variant of PTEN/MMAC1 containing a 9-base pair long deletion.
- 19. (Amended) An oligonucleotide primer for the PCR and the sequencing of exon 1 and/or 5'-untranslated region of the PTEN/MMAC1 gene and its homologues, [characterized by] comprising the following sequences:

PTENex1-401 sense

5'-cccttctactgcctcca -3'

PTENex1 -465 sense

5'- gggagggggtctgagt -3'

PTENex1 ATG sense

5'- atgacagccatcatcaaaga -3'

#### Attachment to Preliminary Amendment dated XXX

## Marked-up Claims 1-45 and 47-52

PTENex1 R antisense

5'- aggtcaagtctaagtcgaatc -3'

20. (Amended) The oligonucleotide primer for PCR and the sequencing of exon 2 of the PTEN/MMAC1 gene and its homologues, [characterized by] comprising the following sequences:

PTENex2F sense

5'- atatttatccaaacattattgctat -3'

PTENex2R antisense

5'- cttactacatcatcatattgttcc -3'

21. (Amended) The oligonucleotide primer for PCR and the sequencing of exon 4, intron 4 and exon 5 of the PTEN/MMAC1 gene and its homologues, [characterized by] comprising the following sequences:

Zoo43sUV sense

5'- tgtgctgagagacattatgac -3'

SPL5 sense

5'- aaatttaattgcagaggt -3'

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## **Attachment to Preliminary Amendment dated XXX**

#### Marked-up Claims 1-45 and 47-52

Zoo44aRV antisense

5'- ttgtctctggtccttacttc -3'

22. (Amended) The oligonucleotide primer for PCR and the sequencing of exon 5 of the PTEN/MMAC1 gene and its homologues, [characterized by] comprising the following sequences:

PTEN se sense

5'- atcttgaccaatggctaagtg -3'

Zoo44aRV antisense

5'- ttgtctctggtccttacttc -3'

23. (Amended) The oligonucleotide primer for PCR and the sequencing of exon 6 of the PTEN/MMAC1 gene and its homologues, [characterized by] comprising the following sequences:

PTENex6F sense

5'- gga gta act att ccc agt cag ag -3'

PTENex6R antisense

 $R: \label{locsloss} $$R: \label{locsloss} $$\Omega = 1.012627 \end{\ensuremath{\mbox{01-09-12}}} $$ amend. wpd$ 

## **Attachment to Preliminary Amendment dated XXX**

#### Marked-up Claims 1-45 and 47-52

5'- gca agt tcc gcc act gaa -3'

24. (Amended) The oligonucleotide primer for PCR and the sequencing of exon 7 of the PTEN/MMAC1 gene and its homologues, [characterized by] comprising the following sequences:

PTENex7F sense

5'- cet cag ttt gtg gtc tgc ca -3'

PTENex7R antisense

5'- c ctt ttt tag cat ctt gtt ctg ttt -3'

25. (Amended) The oligonucleotide primer for PCR and the sequencing of exon 8 of the PTEN/MMAC1 gene and its homologues, [characterized by] comprising the following sequences:

PTENex8F sense

5'- caa aat gtt tca ctt ttg ggt aaa -3'

PTENex8R antisense

5'- taa aat ttg gag aaa agt atc ggt t -3'

#### **Attachment to Preliminary Amendment dated XXX**

# Marked-up Claims 1-45 and 47-52

26. (Amended) The oligonucleotide primer for PCR and the sequencing of exon 9 of the PTEN/MMAC1 gene and its homologues, [characterized by] comprising the following sequences:

PTENex9F sense

5'- gtg aag ctg tac ttc aca aaa ac -3'

PTENex9tga antisense

5'- aaa aaa att cag act ttt gta att tg -3'

- 27. (Amended) The method according to [claims 1 to 17] claim 1, [characterized in that for] wherein the DNA amplification involves a mixture of oligonucleotides [is used] which differ at the 3' region of the oligonucleotide as regards its length by one or more nucleotides or which differ as regards their nucleotide sequence at the 3' end of the oligonucleotide at one or more positions.
- 28. (Amended) The method according to [claims 1 to 17 and 26] claim 1, wherein the oligonucleotides

sense:

5'- cga cgt tgt aaa acg acg gcc agt tgt gct gag aga cat tat gac -3', R:\012627\025\Docs\01-09-12 amend.wpd

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#### Marked-up Claims 1-45 and 47-52

- 5'- cga cgt tgt aaa acg acg gcc agt tgt gct gag aga cat tat -3',
- 5'- cga cgt tgt aaa acg acg gcc agt tgt gct gag aga cat t -3',

#### antisense:

- 5' cag gaa aca get atg act tgt etc tgg tec tta ett e -3',
- 5'- cag gaa aca gct atg act tgt etc tgg tec tta c -3',
- 5'- cag gaa aca gct atg act tgt ctc tgg tcc t -3'

are used for the amplification.

29. (Amended) The method according to [claims 1 to 17 and 26] claim 1, wherein the oligonucleotides

#### sense:

- 5'- cga cgt tgt aaa acg acg gcc agt tgt gct gag aga cat tat gaa -3',
- 5'- cga cgt tgt aaa acg acg gcc agt tgt gct gag aga cat tat gac -3',
- 5'- cga cgt tgt aaa acg acg gcc agt tgt gct gag aga cat tat gag -3',
- 5'- cga cgt tgt aaa acg acg gcc agt tgt gct gag aga cat tat gat -3',

#### antisense:

- 5'- cag gaa aca get atg act tgt etc tgg tee tta ett a -3',
- 5'- cag gaa aca gct atg act tgt ctc tgg tcc tta ctt c -3',
- 5'- cag gaa aca get atg act tgt etc tgg tee tta ett g -3',

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# Marked-up Claims 1-45 and 47-52

5'- cag gaa aca gct atg act tgt ctc tgg tcc tta ctt t -3'

are used for the amplification.

- 30. (Amended) The method according to [claims 1 to 17] claim 1, [characterized in that] wherein DNA sequencing methods are used for the genetic analysis.
- 31. (Amended) The method according to [claims 1 to 17] claim 1, [characterized in that] wherein DNA sequencing techniques are used in the genetic analysis for the PTEN/MMAC1 and/or its pseudogenes and their homologues.
- 32. (Amended) The method of distinguishing the DNA of various species, [characterized in that] wherein at least one hybridization probe pair is used, the melting points of different combinations are determined and compiled for each species into a panel.
- 33. (Amended) The method of distinguishing the DNA of various species, [characterized in that] wherein at least one hybridization probe pair is used and at least one gene segment is amplified, differing hybridization probe pairs hybridize to different gene segments, and the melting points of the different combinations are determined and compiled

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#### Marked-up Claims 1-45 and 47-52

for each species into a panel and/or compared with this panel for the purpose of identification.

- 34. (Amended) The method of distinguishing the DNA of different species according to claim 33, [characterized in that] wherein at least one hybridization probe pair is used and at least one gene segment of at least one species is amplified, differing hybridization probe pairs hybridize to different gene segments of various species, and the melting points of the different combinations are determined and compiled for each species into a panel and/or compared with this panel for the purpose of identification.
- 35. (Amended) The method of distinguishing the DNA of various species according to [claims 33 and 34] claim 33, [characterized in that] wherein at least two hybridization probes of SEQ Nos. 3 to 8 are used, the melting points of different combinations are determined and compiled for each species into a panel.
- 36. (Amended) The method according to [claims 33 and 34] <u>claim 33</u>, [characterized in that] <u>wherein</u> the species differentiation of pig DNA from various other species is made using the hybridization probe pair Al/A2 as the hybridization probe pair.

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## Marked-up Claims 1-45 and 47-52

- 37. (Amended) The method according to [claims 33 and 34] claim 33, [characterized in that] wherein the hybridization probes are used in combinations Cl/C2; A1/B2; A1/A2; C1/A2; B1/B2; B1/A2 for the species differentiation between various species.
- 38. (Amended) LightCycler hybridization probes for exon 5, [characterized by] comprising the sequences:

A1: 5'- tgc ata ttt gtt tca tcc ggg caa att -fluorescein -3'

A2: 5'- LC Red 705 - tta aag gca caa gat ttc tat ggg ga - ph -3'

B1: 5'- tgc ata ttt att aca tcg ggg caa att -fluorescein -3'

B2: 5'- LC Red 640 - aag gca caa gag gcc cta gat ttc ta - ph -3'

C1: 5'-tgc ata ttt gtt aca tcg ggg taa att fluorescein -3'

C2: 5'- LC Red 640 - aag gca caa gag gcc cta gat ttc ta - ph -3'

39. (Amended) LightCycler hybridization probes for exon 6, [characterized by] comprising the sequences

PTENex6FL

5'- tca tct gga tta tag acc agt ggc act - fluorescein -3'

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#### Marked-up Claims 1-45 and 47-52

PTENex6LC 640

5'- LC Red 640 - ttc aca aga tga tgt ttg aaa cta ttc caa- ph -3'

PTENex6F\*

5'- gtg cca ctg gtc tat aat cca gat- fluorescein -3'

PTENex6L\* 705

5'- LC Red 705- ttc ttt aac agg tag cta taa taa tac aca ta- ph -3'

40. (Amended) [The] LightCycler hybridization probes for exon 7, [characterized by] comprising the sequences

PTENex7F\*

5'- taa agg tga aga tat att cct cca att ca - fluorescein -3'

PTENex7L\*640

5'-LC Red 640- acc cac acg acg gga aga caa g - ph -3'

PTENex7 FL

5'-ggtaacggctgagggaactcaaagtac - fluorescein -3'

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# **Attachment to Preliminary Amendment dated XXX**

# Marked-up Claims 1-45 and 47-52

PTENex7 LC (705-labeled)

5'-LC Red 705- tgaacttgtcttcccgtcgtgtgg- ph -3'

41. (Amended) [The] LightCycler hybridization probes for exon 8, [characterized by] comprising the following sequences

PTENex8F\*

5'- tga caa gga ata tct agt act tac ttt aac aaa-fluorescein -3'

PPTENex8L\* 705

5'-LC Red 705 - ctt gac aaa gca aat aaa gac aaa gc- ph -3'

PTENex8 FLU

5' - tgctatcgatttcttgatcacatagacttccatttt - fluorescein -3'

PTENex8 LCR (640-labeled)

5'-LC Red 640- actttttctgaggtttcctctggtcctggtat - ph -3'

42. (Amended) [The] LightCycler hybridization probes for exon 9, [characterized by] comprising the following sequences

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# Marked-up Claims 1-45 and 47-52

PTENex9 FL

5'-aac atc tgg tgt tac aga agt tga act gct- fluorescein -3'

PTENex9 LC 640

5'-LC-640- cct ctg gat ttg acg gct cct cta ct - ph -3'

43. (Amended) Hybridization probe pair AI/A2: specific to PTEN pseudogene pig, [characterized by] comprising

SEQ No. 3 A1: 5'- tgc ata ttt gtt tca tcc ggg caa att -fluorescein -3'

SEQ No. 4 A2: 5'-LC Red 705- tta aag gca caa gat ttc tat ggg ga - ph -3'

44. (Amended) Hybridization probe pair B1/B2: specific to pseudogene man, [characterized by] comprising

SEQ No. 5 B1: 5'- tgc ata ttt att aca tcg ggg caa att -fluorescein -3'

SEQ No. 6 B2: 5'-LC Red 640- aag gca caa gag gcc cta gat ttc ta -ph -3'

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# Marked-up Claims 1-45 and 47-52

45. (Amended) Hybridization probe pair C1/C2: specific to PTEN pseudogene man (C2) and homologue of pig (C1) [characterized by] comprising

SEQ No. 7 - C1: 5'- tgc ata ttt gtt aca tcg ggg taa att - fluorescein -3' SEQ No. 8 - C2: corresponds to probe B2.

- 47. (Amended) DNA sequences of homologues of the PTEN/MMAC1 gene and/or of homologues of the PTEN/MMAC1 pseudogene, which are compiled in the annex under "list of species sequences", which as compared to the PTEN/MMAC1 gene and/or the PTEN/MMAC1 pseudogene comprise genetic variants [such as] comprising base substitutions and/or insertions and/or deletions and are suited for identifying corresponding species.
- 48. (Amended) A kit for carrying out the method according to [claims 1 to 18 and further claims] claim 1, comprising:
  - a) one or more vessels comprising PCR and/or sequencing oligonucleotides binding to highly conserved genes, the oligonucleotides being optionally labeled radioactively or by means of a dye or in another way,
  - b) vessels having further common reagents for DNA amplification and/or DNA analysis, [in particular for DNA sequencing,]

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and

- a vessel containing a control DNA which is suited for testing the oligonucleotides and the reaction conditions.
- 49. (Amended) The kit according to claim 48 [for carrying out the method according to claims 1 to 18 and further claims], comprising:
  - a) one or more vessels with PCR and/or sequencing oligonucleotides [according to claims 19 to 26].
- 50. (Amended) [Kit] The kit for identifying species for carrying out the method according to [claims 1 to 18 and further claims] claim 1, comprising:
  - a) a vessel having an oligonucleotide pair comprising the following sequences:

    5'- cga cgt tgt aaa acg acg gcc agt tgt gct gag aga cat tat gac -3' and 5'- cag
    gaa aca gct atg act tgt ctc tgg tcc tta ctt c -3',
  - b) two vessels with one of the following sequencing oligonucleotides each, these oligonucleotides being optionally labeled radioactively or by means of a dye or in another way:
    - 5'- cag gaa aca gct atg ac -3' and
    - 5'- cga cgt tgt aaa acg acg gcc agt -3',

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# Marked-up Claims 1-45 and 47-52

- c) a vessel containing a control *DNA*, which is suited for testing the oligonucleotides and the reaction conditions.
- 51. (Amended) The kit (Light Cycler Kit) for carrying out the method according to [claims 32 to 37 and further claims] claim 32, comprising
  - a) one or more vessels containing PCR primers and hybridization probes, which bind to highly conserved genes, the hybridization probes being optionally labeled by means of a dye,
  - b) vessels containing further common reagents for DNA amplification and/or DNA analysis, [in particular for the Light Cycler Analyses,]

and

. . A

- a vessel containing a control DNA which is suited for testing the oligonucleotides and the reaction conditions.
- 52. (Amended) The kit (Light Cycler Kit) for carrying out the method according [to claims 32 to 37 and further claims] claim 32, comprising:
  - a) one or more vessels with PCR primers and hybridization probes [according to claims 38 to 42].

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# Method for Identifying Organisms by Means of Comparative Genetic Analysis and Primers and Hybridization Probes for Carrying out this Method

# Description

The present invention relates to a method for the genetic analysis of organisms of different species of animal and/or plant by studying coding and non-coding areas of highly conserved genes or pseudogenes and their homologues with various species of animal and plant.

The known methods serving for simply, readily and precisely determining gene sequences to detect relationships and to identify organisms are based on the use of oligonucleotides which are specific to a species. These oligonucleotides are necessary to provide sufficient genetic material for the subsequent sequence reaction by means of polymerase chain reaction or other methods. An oligonucleotide is usually a short synthetically produced molecule which binds specific gene segment and has a specific sequence complementary to this strand. The drawback of methods based the use of oligonucleotides consists in that gene sequences of different species usually differ strongly. In order to sequence organisms of different species, speciesspecific DNA sequences must, as a rule, be determined by means of methods which are costly and time-consuming, and the corresponding oligonucleotides which bind specifically to these DNA sequences must be synthesized in a second step. These two steps are usually required for each organism to be studied of unknown origin. For a subsequent analysis for determining the identity or the relationship due to the gene sequence the DNA of the organisms to be studied must then be tested with a usually large number of different oligonucleotides. Since usually only oligonucleotides of few species are available, the analysis often fails to be a success in the case of rare species of animal. The analysis is also time-consuming and expensive.

Other methods which utilize e.g. restriction length polymorphisms of the organisms or methods used for amplifying a mixture of short oligonucleotides randomized as regards their sequence (random amplified polymorphic DNA, RAPD) do not result in an accurate and unambiguous analysis of the gene sequence of individual animals. As a result, it is difficult to determine the relationship between organisms by means of these methods.

It is thus the object of this invention to provide a method for the genetic analysis of organisms, which is highly sensitive, supplies reliable results while consuming little time, is also suited for major serial examinations and routine tests, and can optionally also be carried out automatically. The object of this invention also consists in developing means or products for carrying out this method.

The invention is realized according to the claims, the subclaims being preferred variants.

The subject matter of the invention relates to a method by which the gene sequence of a gene area of various species can be determined readily, simply and safely in a reproducible Three preconditions must be met here: The oligonucleotides serving as primers for the amplification of the DNA must bind to areas of the genome which are highly conserved to ensure an amplification of genetic material by means of an identical oligonucleotide pair in all or the greatest possible number of different organisms. oligonucleotides must cover an area which has a great sequence diversity between different species to enable a differentiation. The area which is covered by the oligonucleotides used as primers should be as small as possible to maximize the yields of amplificates and to ensure that copies can also be obtained from strongly degraded DNA in the starting material.

According to the invention this object is achieved by providing a method of determining the identity or the

relationship by comparing coding and non-coding areas of highly conserved genes of pseudogenes and homologues. As a result, it is ensured that a single oligonucleotide pair binds to DNA sequences highly conserved between various species and thus enables a gene segment identical for all species to be amplified. The oligonucleotides comprise one or more gene areas having the greatest possible sequence differences between different species. The determination of the gene sequence of this highly polymorphous gene area in a subsequent reaction step enables the gene sequence to be allocated to a specific species.

Genotyping is made by sequencing or by other methods which are suited for the detection of sequence variants. This comprises genotyping methods assisted by polymerase chain reaction (PCR), such as allele-specific PCR, other genotyping methods using oligonucleotides (e.g. "dot blotting", "Oligonucleotide Ligation Assays" (OLA)), methods using restriction enzymes, analysis of length polymorphisms and "single nucleotide polymorphisms" (SNP), analysis by means of spectroscopic methods such as "matrix-assisted laser desorption/ionization mass spectroscopy" (MALDI), chromatographic methods such as DHPLC for separating strands of differing lengths and sequences and in principle any method available at present or in the future for variant detection, including DNA, RNA and PNA hybridization methods, light cycler technology, TaqMan and molecular beacon technology and the chip technology in all its technological realizations.

The following steps are preferably carried out in the method according to the invention:

- a) DNA isolation: DNA is isolated and purified from blood samples, tissues, hair, foodstuffs and samples containing DNA.
- b) Polymerase chain reaction: The polymerase chain reaction serves for amplifying DNA for the subsequent sequencing reaction. In the polymerase chain reaction, one or more oligonucleotide pairs bind to the DNA to be analyzed (template DNA) of genes which are highly conserved between organisms of various species. In each case, one of the

molecules of an oligonucleotide pair (sense and antisense oligonucleotide) is complementary to one of the two template DNA strands at the 5' or 3' end of a DNA sequence. The binding is oriented such that the synthesis products obtained in an oligonucleotide-controlled polymerase chain reaction using one of the two oligonucleotides each may serve, a matrix for binding the denaturation, as following respectively other oligonucleotide. The oligonucleotide pair flanks the area which shall be copied. These oligonucleotides are extended in accordance with the nucleotide sequence of the template strand by means of polymerase and following the addition of nucleotide building blocks. Binding, extension and denaturation take place at different temperatures and are usually carried out 20 to 35 times in succession so as to multiply exponentially the area covered by the oligonucleotides.

- c) Agarose gel electrophoresis: The DNA fragments are separated from the oligonucleotides in an agarose gel with a voltage being applied, and the band specific to the PCR product is excised using a scalpel and purified.
- d) Sequencing reaction: Another polymerase is added as well as all nucleotide building blocks and special nucleotide building blocks which terminate the chain in the extending reaction. As a result, DNA fragments form which differ in length by one nucleotide each. Areas within the gene are sequenced which as regards their gene sequence are polymorphous between the different species. Therefore, the DNA sequence characteristic of a species can be determined and allocated to a species.
- e) Polyacrylamide gel eletrophoresis: If after concluding the sequencing reactions the differently long DNA strands are separated on a high-resolution polyacrylamide gel in an electric field, shorter DNA strands will migrate more rapidly than longer strands. In the pattern of bands forming, the order from shorter band to the respective band next in length allocated to the corresponding bases A, C, G or T corresponds to the complementary DNA sequence of the template. As a result, the sequence of the DNA strand, which was amplified by means of polymerase chain reaction

beforehand, becomes readable.

f) Comparative analysis of the gene sequence between the animal species and storage of the sequencing data.

For the amplification the method according to the invention preferably uses oligonucleotide pairs which bind to coding or non-coding areas of highly conserved genes or pseudogenes and their homologues, i.e. to genes or pseudogenes and their homologues which show no, or only minor, sequence differences between the individual species. This ensures that an amplificate can be formed in each gene segment to be analyzed of the most differing species by means of a primer pair.

The method according to the invention preferably serves for analyzing areas of the gene or pseudogene and their homologues, which differ as regards their gene sequence between organisms of different species. These may be sequences of coding or non-coding DNA.

The method according to the invention uses for certain batches for the polymerase chain reaction several differently long oligonucleotide pairs in a reaction mixture (multiplex PCR), all of which are partially identical with a starting sequence. Here, the sense or antisense oligonucleotides differ each as regards the nucleotide sequence such that the lengths of some oligonucleotides differ in the 3' region by one or more nucleotides. The use of several oligonucleotides differing in length shall ensure that binding of the oligonucleotides to the template DNA will be possible even if the template DNA differs from some oligonucleotides used as regards the 3' region. The correspondence in the 3' region of the oligonucleotides with the template DNA is essential for the specific amplification and should thus be as precise as possible in this area.

# Example:

#### sense:

- 5'- cga cgt tgt aaa acg acg gcc agt tgt gct gag aga cat tat gac -3',
- 5'- cga cgt tgt aaa acg acg gcc agt tgt gct gag aga cat tat -3',
- 5'- cga cgt tgt aaa acg acg gcc agt tgt gct gag aga cat t -

#### antisense:

- 5'- cag gaa aca gct atg act tgt ctc tgg tcc tta ctt c -3',
- 5'- cag gaa aca gct atg act tgt ctc tgg tcc tta c -3',
- 5'- cag gaa aca gct atg act tgt ctc tgg tcc t -3'.

In another preferred embodiment of the method according to the invention, multiple oligonucleotide pairs are used in a multiplex PCR, which have equal length but differ at one or more positions of the 3' end of the oligonucleotides as regards their nucleotide sequence. The use of a reaction mixture of several oligonucleotides differing as regards the base sequence at the 3' end of the oligonucleotide shall ensure that an amplificate can be formed in the most different species by means of a primer pair.

The method according to the invention preferably analyzes segments of the gene or pseudogene and their homologues which differ as regards their gene sequence between organisms of different species. These may be sequences of coding or non-coding DNA.

The method according to the invention uses for certain batches for the polymerase chain reaction several differently long oligonucleotide pairs in a reaction mixture (multiplex PCR), all of which are partially identical with a starting sequence. Here, the sense or antisense oligonucleotides differ each as regards the nucleotide sequence such that the lengths of some oligonucleotides differ in the 3' region by one or more nucleotides. The use of several oligonucleotides differing in length shall ensure that binding of the oligonucleotides to the template DNA will be possible even if the template DNA differs from some oligonucleotides used as regards the 3' region. The correspondence in the 3' region of the oligonucleotides with the template DNA is essential for the specific amplification and should thus be as precise as possible in this area.

# Example

## sense:

- 5'- cga cgt tgt aaa acg acg gcc agt tgt gct gag aga cat tat gac -3',
- 5'- cga cgt tgt aaa acg acg gcc agt tgt gct gag aga cat tat

- -3',
- 5' cga cgt tgt aaa acg acg gcc agt tgt gct gag aga cat t -3',

#### antisense:

- 5'- cag gaa aca gct atg act tgt ctc tgg tcc tta ctt c -3',
- 5'- cag gaa aca gct atg act tgt ctc tgg tcc tta c -3',
- 5'- cag gaa aca gct atg act tgt ctc tgg tcc t -3'.

In another preferred embodiment, the method according to the invention uses multiple oligonucleotide pairs in a multiplex PCR, which have equal length but differ at one or more positions of the 3' end of the oligonucleotides as regards the nucleotide sequence. The use of a reaction mixture of several oligonucleotides differing as regards the 3' end of the oligonucleotide shall ensure that the oligonucleotides are bound to the template DNA even if the template DNA differs at the 3' binding site of the oligonucleotide from the usually used oligonucleotide. For this purpose, a mixture of different oligonucleotides which have all conceivable nucleotide sequences at their 3' end is provided for the amplification.

#### Example:

#### sense:

- 5'- cga cgt tgt aaa acg acg gcc agt tgt gct gag aga cat tat gaa -3',
- 5'- cga cgt tgt aaa acg acg gcc agt tgt gct gag aga cat tat gac -3',
- 5'- cga cgt tgt aaa acg acg gcc agt tgt gct gag aga cat tat gag -3',
- 5'- cga cgt tgt aaa acg acg gcc agt tgt gct gag aga cat tat gat -3',

# antisense:

- 5'- cag gaa aca gct atg act tgt ctc tgg tcc tta ctt a -3',
- 5'- cag gaa aca gct atg act tgt ctc tgg tcc tta ctt c -3',
- 5'- cag gaa aca gct atg act tgt ctc tgg tcc tta ctt g -3',
- 5'- cag gaa aca gct atg act tgt ctc tgg tcc tta ctt t -3'. When suitable oligonucleotide sequences are selected, the method according to the invention pays attention to the fact that as many oligonucleotides as possible correspond at their 3' end with the first nucleotide of the codon which codes for

the highly conserved amino acid. It follows from theoretical considerations and also on the basis of observations that every second nucleotide has within a codon a degree of correspondence between organisms of differing species higher than that of the first or third nucleotide of the codon. Since the amplification will usually only function if along with other preconditions the nucleotide located at the 3' end of the oligonucleotide is exactly complementary to the opposite nucleotide of the template DNA strand - i.e. A faces T or G faces C -, the sequence of the oligonucleotides employed is chosen such that the last nucleotide at the 3' end of the oligonucleotide binds, if possible, to the most highly conserved nucleotide of the codon coding for an amino acid.

The method according to the invention preferably uses for different segments of the gene, of the pseudogenes and its homologues a single oligonucleotide pair each, which binds to the highly conserved tumor suppressor gene PTEN/MMAC1, its pseudogene and their homologues in different species, namely in the area of the gene which shows major correspondence between the species [Steck et al., 1997]. This area concerns the entire coding sequence of the gene, pseudogene and its homologues as well as exon-intron transitions and 5'- and 3'-untranslated regions of the gene, pseudogene and their homologues.

The areas amplified by means of the above described methods comprise sequence segments which have more or less great sequence differences between the individual species. This applies in particular to intron regions and specifically to intron 4. The oligonucleotide pair used for the amplification may bind to exons 4 and 5, since the PTEN/MMAC1 gene, pseudogene and their homologues show no, or only slight, differences between the species in exon 4 and exon 5. The intron region 4 covered by both oligonucleotides, which in contrast to the exon regions 4 and 5 between the species has considerably greater sequence differences, is amplified. Thereafter, part of this intron region is sequenced preferably by means of a sequencing reaction (see figure 1).

By means of comparative analyses, it is possible to determine due to the correspondence of intron sequences whether different samples belong to an identical species. It is also possible to determine the relationship of different species by means of comparative analysis of an intron region comprising only some to several hundred bases of PTEN/MMAC1 gene and its homologues due to the similarity of the sequences. The general rule is that for the differentiation of closely related species the intron segments which must be studied have to be longer than those of distantly related species. This procedure described for exons 4/5 and intron 4 may also be applied to all of the other introns enclosed by exons. It also applies to pure exon regions, pseudogenes and the 5'- and 3'-untranslated regions as well as their homologues, in these cases the sequence differences between the individual species being less than in the intron regions.

In a preferred embodiment of the method according to the invention, highly conserved pseudogenes and their homologues are studied in various organisms and used for determining the species-specific gene sequence. These pseudogenes and their homologues have the advantage that they are also highly conserved as regards their nucleotide sequence and allow an amplification in certain species. However, the sequence differs in some areas of the pseudogene and its homologues between different species, so that pseudogenes and their homologues can be used for the species-specific characterization of organisms. Since the intron regions lack, they - like pure exon regions and the 5'- and 3'-untranslated regions - are suitable for the analysis, in particular of DNA degraded due to environmental influences, because of the small size.

An advantageous execution of the method utilizes the deletion identified by inventors and having a length of 9 base pairs in a PCR product (see figure 3 and Example 2), which corresponds to a gene sequence variant of PTEN/MMAC1 and was amplified from DNA of pig cells. This deletion having a

length of 9 base pairs is typically found in domestic pigs and all examined wild boars and as for the rest in no other examined species. In a variant of an embodiment of the invention this difference in length serves for proving pig meat in foodstuffs. This variant in length is genotyped by sequencing or by other methods suitable for detecting this deletion. They comprise PCR-assisted genotyping methods such PCR by means of species-specific oligonucleotides, hybridization techniques such as the light cycler technology or other genotyping methods using restriction enzymes, and in principle any method available at present or in the future for detecting variants, including the chip technology and all their technological realizations. Deletions and insertions, as found in different species in intron 4, exon 8 in the 5'-untranslated region, can also be used correspondingly for identifying species (see annex: "List of species sequences").

The method according to the invention was made with DNA from different species as a model system and the gene sequence in the area of two segments in intron 4 of the PTEN/MMAC1 gene and its homologues was determined. It was possible to amplify species with only one oligonucleotide pair. sequencing reaction with only one oligonucleotide and in the subsequent analytical polyacrylamide gel electrophoreses it turned out that all investigated species differ as regards the nucleotide sequence in the intron region (see annex: "List of species sequences"). Since the PTEN/MMAC1 tumor suppressor gene and its pseudogene and their homologues are conserved, the method can in other species also amplify successfully the corresponding gene sequences of organisms and possibly plants by means of an oligonucleotide pair. The method according to the invention is thus suited to determine the identity and relationship of various organisms. A data library was established which can be used for identifying different species and humans. It comprises PCR primers, sequencing primers and hybridization probes as well as sequences of the coding and non-coding areas including select highly variant intron regions, of exon regions, of the 5'-untranslated region of the gene and its homologues as well the pseudogene and its homologues from the most differing vertebrates (see annex: "List of species sequences"). On the one hand, the method according to the invention is suited to determine readily, simply and safely in a reproducible manner the relationship in certain species which are clearly classified (phylogenetic analyses). On the other hand, it is possible to determine by a comparison with gene sequences which can clearly be allocated to a species, the identity of tissue samples, blood samples and foodstuffs and all samples which contain DNA and are of unknown origin. Therefore, the method is also suited for applications in forensic medicine. Since in a preferred embodiment of the method according to the invention DNA is used as the starting organic samples (e.g. blood, saliva, material, residues) can clearly be allocated to human or animal origin because of their gene sequence. Due to the possibility of comparing the collected DNA sequence with an already established data library with DNA sequences of known species it is possible to make statements on the species. If the gene sequence is unknown, it is possible due to sequence similarities to make statements on the relationship which the species to be studied has with a comparative DNA.

An advantageous embodiment of the invention utilizes hybridization probes for distinguishing the DNAs of various species among one another. This method uses differing ways for distinguishing the species on the basis of the LightCycler analysis system (company of Roche Molecular Biochemicals) using hybridization probes (patents WO 97/46714; WO 97/39008).

The LightCycler analysis system makes possible the amplification, detection and specific analysis of DNA of differing species and their differentiation within the shortest possible time and with moderate expenditure. The LightCycler is a micro-volume fluorimeter having a thermocycler combining rapid thermocycling with real-time fluorescence observation during the PCR (Wittwer et al., 1997a). By means of this technology, the time for the

amplification and the detection of nucleic acids is reduced from about 5 hours to about 30 minutes. For a specific detection during the PCR reaction, the synthesis can be observed on the basis of the fluorescence resonance energy transfer (FRET) via two adjacent hybridization probes labeled with fluorescent dyes. Here, one probe is labeled at its 3' end with a donor fluorophore (usually fluorescein) and the adjacent probe is labeled at its 5' end with an acceptor fluorophore. During FRET, the donor dye is excited by an external light source and emits light which is absorbed by the acceptor fluororphore. The latter in turn emits light having another wavelength which is measured specifically. This FRET can only be made if both probes hybridize side by side within the amplification pair at a distance of about 1-5 bp, on the target DNA. Here, the sequence of the probes is selected such that it is complementary to the target area [Wittwer et al. 1997b; Lay, 1997, Bernard, 1998; Ririe, 1997; Bernard, 1999; Nauck, 1999a; Nauck, 1999b, Kreuzer, 1999; Kyger, 1998; Mangasser-Stephan, 1999; Aslandis, 1999].

In order to detect the specific amplification product and to differentiate differences in the target sequence, the LightCycler provides the possibility of carrying out melting point analyses. The melting point analysis is based on the fact that two complementary DNA strands are separated at a characteristic temperature, the melting temperature, into two individual strands. This melting temperature depends on the base composition in which sequences rich in GC have a higher melting point than sequences with predominantly AT bases. If a melting point analysis is carried out by removing more or less well complementary hybridization probes from the template DNA under certain temperature conditions, there is a melting profile characteristic of the fitting between probe and target sequence to be analyzed in the form of fluorescence intensity measurement. As a result, the changes in the target sequence can be detected. If the sequences are fully complementary to one another, the probes will fully hybridize. If the target sequence contains changes in its base sequence, the melting point of the probes is lowered correspondingly. Due to a continuous detection of the fluorescence up to the melting of the probes, a melting curve can be prepared for each sample in the form of the fluorescence intensity as a function of the temperature. A comparison of the melting peaks made by the first negative derivative of the fluorescence to the temperature (-dF/dt vs T) enables a verification and differentiation of various DNAs. Fragments having a lower or higher melting temperature can be distinguished clearly.

In particularly preferred variants of embodiments of the invention, oligonucleotide pairs were found for the polymerase chain reaction (PCR) which bind to the highly conserved tumor suppressor gene PTEN/MMAC1, its pseudogene and their homologues in areas which show large correspondence between the species. These are usually exon regions or untranslated areas of the 3' and 5' ends of the gene, its pseudogene and their homologues.

These oligonucleotide pairs allow to amplify small gene segments containing areas which have differing base sequences between the individual species, including base substitutions, base deletions and base insertions. Examples 4, 5 and 6 use a 9-base pair deletion of the pig homologue of the PTEN/MMAC1 pseudogene and numerous further sequence variants of the gene, pseudogene and their homologues in various species to produce by means of specific hybridization probes differing melting profiles with differing species, which clearly distinguish the species from one another. This preferred area of Examples 4 to 6 relates exclusively to exon 5 of the gene, pseudogene and their homologues. The PCR primers used for the amplification of the corresponding gene segment read as follows:

Sense primer: PTEN se 5'- atc ttg acc aat ggc taa gtg -3' Antisense primer: Zoo44aRV 5'- ttgt ctc tgg tcc tta ctt c -3'

Hybridization probes according to the invention were selected on the one hand with the target area of the 9-base pair deletion of the pig pseudogene. Probe A1/A2 enables a distinction of the pig DNA from all of the other species.

Since by the above PCR primer pair, it is not only the pseudogene but also the gene area of exon 5 of the pig homologue of PTEN/MMAC1 that is amplified, another probe pair was provided with the complementary sequence of the pig homologue. This is probe C1/C2.

Since the different species in this gene area show minor sequence differences which should be used for a detailed differentiation among one another, a third probe pair which corresponds to the sequence of the human pseudogene in the selected area was constructed (probe B1/B2).

According to the invention, the probes A1/A2 are concerned: specific to PTEN pseudogene pig:

A1: 5'- tgc ata ttt gtt tca tcc ggg caa att - fluorescein -3'

A2: 5'- LC Red 705 - tta aag gca caa gat ttc tat ggg ga - ph

Probes B1/B2: specific to PTEN pseudogene man:

B1: 5'- tgc ata ttt att aca tcg ggg caa att - fluorescein -3'

B2: 5'- LC Red 640 - aag gca caa gag gcc cta gat ttc ta - ph

Probes C1/C2: specific to PTEN homologue pig:

C1: 5'- tgc ata ttt gtt aca tcg ggg taa att - fluorescein

C2: corresponds to probe B2

The positions of probes A1, A2, B1, B2, C1 and C2 in exon 5 are shown in figure 4.

The separate use of these three probe pairs and the use of the probes in various possible combinations (1 donor dye + 1 acceptor dye) yields for each individual species a characteristic panel of differing melting points enabling a clear distinction between the species (see figures 5 and 7). The use of these hybridization probe combinations also permits to carry out studies in reaction mixtures of two or more different species.

A parallel analysis/detection of the fluorescence of two different wavelengths (640 nm and 705 nm) is possible in a multiplex reaction with two different probe pairs such that the respective donor probes are labeled differently. The acceptor probes may be identical or differ as regards their sequence. Having concluded the melting point analysis, a melting point specific to the corresponding probe and the target sequence covered by it is obtained for each of the two wavelengths.

The alternative combination from two different donor probes and one acceptor probe of one wavelength yields for a reaction mixture of two different species having minor sequence differences in the target area two melting points within one wavelength.

In general, the following general multiplex reaction batches are possible which are characterized in that at least one hybridization probe pair is used and at least one gene segment is amplified, differing hybridization probe pairs hybridize to differing gene segments, and the melting points of the different combinations are determined and compiled for each species into a panel or used for the identification. These general multiplex reaction batches can also be characterized in that not only at least one hybridization probe pair is used and not only at least one gene segment is amplified but also DNA of at least one species is used and thus different hybridization probe pairs hybridize to differing gene segments of different species, and the melting points of the different combinations are determined and compiled for each species into a panel or used for the identification.

On this basis, the following analytical approaches are possible, melting point overlaps having to be avoided.

- a) Analysis of an unknown species sample with two different donor probes and one acceptor probe of one wavelength or of differing wavelengths,
- b) Analysis of two or more unknown species samples with two different donor probes and one acceptor probes of one wavelength or of different wavelengths,
- c) analysis of an unknown species sample with a mixture of two or more donor probes and acceptor probes each,
- d) analysis of a mixture of unknown species samples with one donor probe and one acceptor probe,
- e) analysis of a mixture of unknown species samples with two or more different donor probes and two or more acceptor probes of one or more different wavelengths.

This procedure can be applied to all highly conserved segments of the PTEN/MMAC1 gene, pseudogene and their homologues and also to other highly conserved genes when species shall be distinguished.

Select primers for the amplification are defined in claims 18 to 25 and select hybridization probe sequences for the LightCycler application are defined in claims 34 to 42 for different exons of the PTEN/MMAC1 gene, pseudogene and their homologues. Due to the high conservation of the gene in the evolution, primers and hybridzation probes are also possible in all of the other conceivable areas of the gene, pseudogene and their homologues which can be used according to the above described principle for distinguishing species.

These probes and/or primers can be combined as desired in accordance with the multiplex principle with the aim of differentiating species.

The general rule is that all of the described methods and all methods conceivable at present and in the future for analyzing the DNA sequence variants can be applied to both the sense strand and antisense strand. This equivalence principle also applies to all PCR, sequence primer and hybridization probes which are described in this patent

application and the described gene sequences of the individual species of animal.

The following examples shall explain the invention in more detail.

# Example 1

In this experiment, the sequence diversity between human DNA and elephant DNA is determined. The arrangement shown in figure 1 was chosen. The comparative sequence analysis was carried out by determining the species-specific sequences of intron 4 of the tumor suppressor gene PTEN/MMAC1 and its homologues. An oligonucleotide pair which is specific to areas within exons 4 and 5 of the DNA sequence of this gene for mice was chosen and subsequently synthesized (company of Amersham Pharmacia).

Oligonucleotide 1: 5'- cga cgt tgt aaa acg acg gcc agt tgt gct gag aga cat tat gac -3'
Oligonucleotide 2: 5'- cag gaa aca gct atg act tgt ctc tgg tcc tta ctt c -3'

The oligonucleotides for the polymerase chain reaction were constructed such that they have one sequence each at their 5' end (underlined) which is complementary to the two oligonucleotide used for the sequencing reaction. As a result, it is ensured that the specific oligonucleotide parts bind in each case specifically to the template DNA (not underlined) but the oligonucleotides used for sequencing may differ from those used for the polymerase chain reaction, which adds quite generally to the quality of the sequencing reaction. The specific portion of the oligonucleotides binds in each case to exon 4 and exon 5 of the PTEN/MMAC1 tumor suppressor gene and its homologues.

A veterinarian provided 3 ml of residual blood of an African elephant resulting from a routine operation. The DNAs from elephant blood and from human blood were isolated by means of the QIAgen kit (QIAGen company) in accordance with the protocols from the manufacturer and stored at -20°C until they were used.

In order to generate sufficient amounts of DNA for the sequencing reaction, a polymerase chain reaction (PCR) was carried out. 3 sample batches were provided and the following

volumes and final concentrations or amounts of substrates and units of polymerase were used: reaction vessel 1: elephant DNA 1  $\mu$ l (50 ng); reaction vessel 2: human DNA 1  $\mu$ l (50 ng); reaction vessel 3: no DNA but 1  $\mu$ l distilled water instead (negative control). All sample batches were provided with the following: 14.35  $\mu$ l distilled water; dNTPs (Promega company): 4  $\mu$ l (200  $\mu$ l); MgCl<sub>2</sub> (InViTek): 1  $\mu$ l (2 mM); 10 x buffer. (InViTek) consisting of 160 mM (NH<sub>4</sub>)<sub>2</sub>SO<sub>4</sub>, 500 mM Tris-HCl, pH 8.8, 0.1 % Tween 20: 2.5  $\mu$ l; oligonucleotides 1 and 2: 1  $\mu$ l (0.2  $\mu$ M); Tag polymerase (InViTek): 0.15  $\mu$ l (0.75 units). All in all, 35 cycles were carried out in a Perkin Elmer Thermocycler 9600, denaturing taking place at 94°C for 50 seconds each, the oligonucleotides binding to the DNA strands at 53°C for 50 second, and extension taking place at 72°C for 60 seconds (one second longer per cycle). Before the first cycle was started, denaturing was carried out at 94°C for 3 minutes, and a last extension phase of 10 minutes at 72°C was carried out after the last cycle.

In order to check the success of the PCR, to isolate the amplified DNA and check the negative control, an agarose gel electrophoresis was carried out: A 0.8 % agarose gel was produced by adding ethidium bromide, and 1 x TAE buffer was added. 13  $\mu$ l PCR product each was added by pipetting to prepared sample bags or pockets to the gel matrix by adding 2  $\mu$ l dye and a voltage of 100 V was applied. Electrophoresis was stopped after 20 to 30 minutes, the DNA bands were made visible under a U.V. lamp and thereafter excised using a scalpel. The purification of the PCR product was possible by placing the excised bands on MicroSpin columns (Amersham Pharmacia) and centrifugation at 1020 g for 10 minutes. The eluate was diluted depending on the strength of the band on the agarose gel with up to 30  $\mu$ l distilled water.

The purified PCR products were subjected to a cycle sequencing reaction using oligonucleotides having the sequence 5' - Cy-5- cag gaa aca gct atg ac -3'. The oligonucleotides were labeled at the 5' end with the dye Cy-5. 4 reaction vessels (A, C, G, T) were provided per PCR

product, and the following volumes and concentrations were chosen: 3  $\mu$ l PCR product each per A, C, G, T; 1  $\mu$ l each per A, C, G, T reagent (Amersham Pharmacia) consisting of Tris-HCl (pH 9.5), MqCl<sub>2</sub>, Tween 20, Nonidet P-40, 2-mercaptoethanol, dATP, dCTP, 7-deaza-dGTP, dTTP, thermoresistant pyrophosphatase and thermo sequenase DNA polymerase, reaction vessels A, C, G and T containing correspondingly ddATP, ddCTG, ddGTP and ddTTP. Thereafter, 1  $\mu$ l oligonucleotide of the above mentioned sequence  $(0.5 \mu M)$  is added. The following reaction conditions were chosen: All in all, 25 cycles were carried out in a Perkin Elmer Thermocycler 9600 (Perkin Elmer company), denaturing taking place at 94°C for 20 seconds each, the oligonucleotides binding to the amplificate at 54°C for 30 seconds, and the DNA strands being extended while raising the temperature to the denaturing temperature. Prior to the commencement of the first cycle, denaturing was carried out at 94°C for 3 minutes and 30 seconds, and the last extension phase of 5 minutes was carried out at 72°C after the last cycle.

In order to make visible the sequence, the products were separated on a polyacrylamide gel in an electric field. For this purpose, the automatic laser fluorescence detection system A.L.F. express from the company of Amersham Pharmacia was chosen. The gel matrix was composed of 16.8 g urea (Gibco BRL), 5.2 ml 50 % long ranger gel solution (FMC company) and 4 ml 10 x TBE (Gibco BRL), which were diluted using distilled water to give 40 ml. The gel was polymerized by adding 140  $\mu$ l 10 % APS (Merck company) and 20  $\mu$ l TEMED (Serva company). 5  $\mu$ l formamide loading dye were added to each reaction vessel A, C, G, T, and added by pipetting to the prepared sample pockets. The following electrophoresis conditions were chosen: 1000 V, 40 mA, 40 W.

Following the gel electrophoresis, the sequences were generated by the A.L.F.express system (Amersham Pharmacia) and could then be compared with one another. Figure 2 shows the determined nucleotide sequence differences. A comparison of the sequences between elephant DNA and human DNA in the exon region yielded a great correspondence and served as a

control for the PTEN/MMAC1 specificity, whereas the intron region differed greatly.

# Example 2

In this experiment, the sequence of a pig liver provided by a butcher's is compared with already available DNA of a pig. Beef salami served as a control. DNAs were obtained from 50 to 60 mg pig liver and 50 to 60 mg beef salami using the QIAgen kit (QIAGen) and purified. For the comparative analysis, the procedure carried out in Example 1 was chosen using oligonucleotides 1 and 2 for the polymerase chain reaction. A band having a length of about 300 bp was excised from the agarose gel for the sequence analysis. This band corresponds in length to the PTEN/MMAC1 pseudogene. After purifying the excised amplificate, Cy-5-labeled oligonucleotides with the sequence 5'- Cy-5- cag gaa aca gct atg ac - 3' were sequenced. The comparative analysis of pig liver from a butcher's with pig DNA already sequenced in this area resulted in a full correspondence. The control DNA (beef salami) has a sequence markedly differing from the pig DNA (see figure 3).

# Example 3

The PCR and sequencing of exons 1 to 9 is carried out with the primers described in claims 19 to 26 with otherwise analogous reaction control. The corresponding sequences of the studied species are listed in the annex under "List of species sequences".

# Example 4

This experiment is to show the species differentiation between pig DNA and human DNA using various combinations of hybridization probe pairs. The comparative analysis was made by determining the melting points of the hybridization probe pairs C1+B2, A1+B2, A1+A2, and C1+A2 in pig DNA and human DNA. The probes were chosen correspondingly and synthesized

(Tib Mol Biol, Berlin. Probes:

- A1: 5'- tgc ata ttt gtt tca tcc ggg caa att fluorescein -3'
- A2: 5'- LC Red 705 tta aag gca caa gat ttc tat ggg ga ph 3'
- B1: 5'- tgc ata ttt att aca tcg ggg caa att fluorescein -
- B2: 5' LC Red 640 aag gca caa gag gcc cta gat ttc ta ph -3'
- C1: 5'- tgc ata ttt gtt aca tcg ggg taa att fluorescein -
- C2: 5'- LC Red 640  $\stackrel{\circ}{-}$  aag gca caa gag gcc cta gat ttc ta ph -3'

A precondition for a probe hybridization is the amplification of the target area with a suitable primer pair. For this purpose, the following primer pair was chosen and synthesized:

Sense primer: PTEN se 5'- atc ttg acc aat ggc taa gtg -3'
(Tib Mol Biol)

Antisense primer: Zoo44aRV 5'- ttgt ctc tgg tcc tta ctt c-3' (Amersham Pharmacia)

Both primers bind to areas of the exon 5 of the PTEN/MMAC1 gene, pseudogene and their homologues, which comprises the target regions of about 172 base pairs.

The DNAs were isolated by means of the Quiagen kit (QIAGen) from the blood of pigs and humans in accordance with the protocols of the manufacturer and stored at -20°C until they were used.

The real time PCR with hybridization probes and subsequent melting point analysis was carried out as follows: 3 rows with 4 sample batches each (glass capillary cuvette (Roche)) were provided in a cooled pipetting block (Roche). In row 1,

2  $\mu$ l (50 ng) pig DNA are supplied in each batch, 2  $\mu$ l (50 ng) human DNA are supplied to each batch of the 2<sup>nd</sup> row, and 2  $\mu$ l distilled water are supplied in row 3 for the negative controls. The hybridization probes were used for the corresponding batches in the following combinations: 1<sup>st</sup> batch of each row: A1, 1  $\mu$ l (0.1  $\mu$ M) and A2, 2  $\mu$ l (0.2  $\mu$ M); 2<sup>nd</sup> batch of each row: C1, 1  $\mu$ l (0.1  $\mu$ M) and B2, 2  $\mu$ l (0.2  $\mu$ M); 3<sup>rd</sup> batch of each row: A1, 1  $\mu$ l (0.1  $\mu$ M) and B2, 2  $\mu$ l (0.2  $\mu$ M); 4<sup>th</sup> batch of each row: C1, 1  $\mu$ l (0.1  $\mu$ M) and B2, 2  $\mu$ l (0.2  $\mu$ M).

The following volumes and final concentrations or amounts of substrates and units of polymerase were used in all sample batches: oligonucleotides PTEN se and Zoo44aRV 2  $\mu$ l (10  $\mu$ M);

batches: oligonucleotides PTEN se and Zoo44aRV 2  $\mu$ l (10  $\mu$ M) each; MgCl<sub>2</sub> (Roche Molecular Diagnostics): 2.4  $\mu$ l (4 mM); LightCycler DNA Master Hybridization Probes (Roche Molecular Diagnostics): 2  $\mu$ l of a stock solution concentrated by 10 times from Taq DNA polymerase, reaction buffer, dNTP mixture and 10 mM MgCl<sub>2</sub>. The final MgCl<sub>2</sub> concentration in the total reaction batch of 20  $\mu$ l is 5 mM. After fully loading all reagents, the glass capillary cuvettes are closed, centrifuged at 2000 g for 1 minute and inserted in the reaction carrousel of the LightCycler provided for the capillaries.

All in all, 45 cycles are carried out in the LightCycler analysis system as follows: denaturation at 95°C for 1 second, binding of the oligonucleotides and probes at 54°C for 10 seconds and extension of the DNA strands at 72°C for 5 seconds. Denaturation was carried out at 95°C for 30 seconds before the first cycle started. The preliminary heating rate is programmed from denaturation to binding to 20°C/second, from binding to the extension of the DNA strands to 20°C/second and for the step of extension up to the denaturation to 20°C/second. The fluorescence resulting from the FRET of the probes binding complementarily side by side, is measured for observing the PCR at the end of the binding phase in each cycle. Probes which cannot bind fully to the target DNA due to sequence differences, since their binding temperature is below that of the oligonucleotides, cannot yet

be detected at that stage of fluorescence measurement.

After the complete amplification, a melting curve was finally recorded as follows: denaturation of the amplification products at 95°C for 5 seconds, cooling to 30°C with a preliminary heating rate of 20°C/second, holding of this temperature for 15 seconds and subsequent slow heating of 0.2°C/second up to 95°C. The fluorescence of the bound hybridization probes is recorded continuously up to the respective melting/dissociation during the slow temperature increase. A melting curve is recorded for each sample by recording the fluorescence signal as a function of the temperature. By forming the first negative derivative of the fluorescence against the temperature, the melting curves are converted into melting peaks.

After balancing the fluorescence signals at 640 nm and 705 nm against the standard curves prepared with a colorcompensation kit (Roche Molecular Diagnostics) for the respective wavelength, the melting points of pig DNA could be compared with those of the human DNA for the various probe combinations. In this connection, 2 melting points each could be recorded for the pig DNA for each probe pair, and one could be recorded each for the pseudogene and one for the gene. All melting points of the pig DNA differed from those of the human DNA (figure 5).

#### Example 5

In this experiment, the species differentiation of pig DNA from various other species of animal is shown by means of a single hybridization probe pair. For this purpose, DNAs from cattle, sheep, dwarf goat, chicken and turkey were used for the analysis (gene sequences in the annex "list of species sequences" under "sequences intron 4 exon 5").

The comparative analysis was carried out by determining the melting points of a hybridization probe pair whose sequences are specific to the pseudogene of pigs in the area of the 9 base pair deletion (A1/A2) and which was already used in Example 4. The primers used in Example 4 were employed for

the amplification. The DNA was collected from the bloods of said species by means of the QIAGen kit (QIAGen) and purified. For the real-time PCR a sample batch having 2  $\mu$ l (50 ng) of the corresponding DNA each is supplied for each species and a sample batch with 2  $\mu$ l distilled water is supplied for a negative control. Furthermore, 1  $\mu$ l (1  $\mu$ M) of the probe A1 and 2  $\mu$ l (0.2  $\mu$ M) of probe A2 were added by pipetting to each sample batch. To pipette further reagents the procedure of Example 1 was chosen as regards their volumes and concentrations and for the further reaction steps at the LightCycler.

A comparison of the melting points of pig DNA with those of the other animal species shows a clear distinction between pig DNA and those of other animal species (figure 6).

# Example 6

In this experiment, the species differentiation between various animal species is shown by way of the following species with hybridization probes of Example 4:

Pig, deer, dog, Indian elephant, trout, quail, duck, goitred gazelle, mouse, guinea pig (gene sequences in the annex "list of species sequences" under "sequences intron4 exon5")

A comparative analysis was carried out by determining the melting points from various combinations of the hybridization probes from Example 4 in the combinations C1+B2, A1+B2, A1+A2, C1+A2, B1+B2 and B1+A2 and compiling a panel for each species of animal.

The DNA was obtained from the bloods of said species using the QIAGen kit (QIAGen) and purified. The primers listed in Example 4 were used for the amplification. For the further course of the experiment and the subsequent melting point analysis, the procedure in Example 4 was chosen, complemented by the additional probe combinations and greater numbers of species.

A panel of their melting points was compiled for the

corresponding probe combinations for each species (figure 7). A comparison of the panels of the different species yields a difference of each species from the others results as regards at least one melting point.

Figure 8 shows the mean value curves which were determined for selected probe combinations (C1+C2/A1+A2/B1+A2 and A1+C2 from 15 pig DNAs each and C1+C2/A1+A2/B1+A2 and C1+A2 from 15 human DNAs each) with the corresponding standard deviations. The experiment control corresponds to Example 4.

Furthermore, the sequences of the investigated species for exons 1-9 and the 5'-untranslated region of the PTEN/MMAC1 gene, pseudogene and their homologues are listed in the annex under "list of species sequences".

These examples for various probe combination panels can be enlarged by any number of probes and primers for the described gene PTEN/MMAC1, pseudogene and their homologues, but also for all of the other genes, and applied. A linear increase of the primers and probe combinations in the entire genome yields an exponential growth of differentiation possibilities of the most varying species among one another. By using sequence-specific probes and several and/or any number of probe combinations, a species can be distinguished from all of the other species by its very specific melting point panel.

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#### Annex

# List - figures 1 - 8:

Figure 1: diagram of the method for identifying organisms according to Example 1

- Figure 2: comparison of the determined sequences from man and African elephant according to Example 1
- Figure 3: DNA sequence comparison of a pig with purchased pig liver and beef salami according to Example 2. The differences between the nucleotide sequences are shown. The 9-base pair long deletion (nucleotides 216-224) is striking in pig / pig liver.
- Figure 4: positions of the hybridization probes in exon 5 of the PTEN gene, pseudogene and their homologues
- Figure 5: Example 4 melting point panels of pig and man
- Figure 6: Example 5 melting points of the probe combination A1 + A2 in pig as compared to various species
- Figure 7: Example 6 melting point panel of various species
- Figure 8: standard deviations of select probes for pig (HS) and man (WT)

#### Claims

- 1. A method of identifying organisms by comparative genetic analysis, characterized in that coding and/or non-coding areas and/or functionally significant areas of highly conserved genes and/or their homologous genes and/or their cDNA copies and/or their pseudogenes are amplified using PCR and are subsequently genotyped and analyzed.
- 2. The method according to claim 1, characterized in that one primer pair each is used for each specific segment of the highly conserved gene, which is located in the highly conserved exon region and/or non-coding areas and/or functionally significant areas and/or in the 5'-or 3'-untranslated area of the gene and binds in as many studied species DNAs as possible, preferable in all studied species DNAs, and enables the amplification of the corresponding gene area.
- 3. The method according to claims 1 and 2, characterized in that coding and/or non-coding areas located between the primers and being either highly variant intron regions and/or variant exon regions or 5'- or 3'-untranslated areas of the gene, are analyzed as regards their sequence and identified by comparison with the species-specific sequence variants.
- 4. The method according to claims 1 to 3, characterized in that either the sense strand or the antisense strand of any species DNA or also their PCR copies are used for the identification.
- 5. The method according to claims 1 to 4, characterized in that preferably animals are identified.
- 6. The method according to claims 1 to 4, characterized in that preferably vertebrates are identified.
- 7. The method according to claims 1 to 4, characterized in

that preferably mammals are identified.

- 8. The method according to claims 1 to 4, characterized in that preferably plants are identified.
- 9. The method according to claims 1 to 4, characterized in that genotying is carried out by DNA sequencing, any hybridization methods, restriction fragment length analyses, chromatographic methods, spectroscopic and in particular mass-spectroscopic methods, allele-specific PCR or by other methods suitable for detecting DNA sequence variants.
- 10. The method according to claims 1 to 4, characterized in that exon and/or intron areas as well as functionally significant areas of the highly conserved tumor suppressor gene PTEN/MMAC1 and its homologues are used for amplification and subsequent genetic analysis.
- 11. The method according to claims 1 to 4, wherein cDNA copies of the PTEN/MMAC1 gene and its homologues are used for the genetic analysis.
- 12. The method according to claims 1 to 4, wherein pseudogenes or segments of pseudogenes of the PTEN/MMAC1 gene and its homologues are used for the genetic analysis.
- 13. The method according to claims 1 to 4, characterized in that preferably exons arranged side by side of the PTEN/MMAC1 gene and its homologues and/or the parts of the introns following the exons are analyzed genetically.
- 14. The method according to claims 1 to 4, characterized in that the exon regions 1 and 2 and/or 3 and 4 and/or 4 and 5 and/or 5 and 6 and/or 6 and 7 and/or 7 and 8 and/or 8 and 9 with the enclosed intron regions 1 and/or 2 and/or 3 and/or 4 and/or 5 and/or 6 and/or 7 and/or 8

as well as the 5'- and 3'-untranslated regions of the PTEN/MMAC1 gene and their homologues are used for the genetic analysis.

- 15. The method according to claims 1 to 4, characterized by selecting areas of highly conserved genes and/or pseudogenes and their homologues, constructing suitable oligonucleotides as primers which bind to the corresponding complementary coding and/or non-coding areas and/or functionally significant areas, amplifying them by means of a suitable technique and comparatively analyzing the sequence of the corresponding coding and/or non-coding area of various species by genetic analysis.
- 16. The method according to claim 15, characterized in that areas of the PTEN/MMAC1 gene and/or the pseudogene and their homologues are selected.
- 17. The method according to claims 15 and 16, characterized in that differing sequence segments of each individual exon, intron or untranslated region of the PTEN/MMAC1 gene and their homologues or the corresponding cDNA are selected.
- 18. The method according to claims 1 to 17, characterized in that genotyping of pig DNA which is obtained preferably from foodstuffs, is carried out on the basis of the gene sequence variant of PTEN/MMAC1 containing a 9-base pair long deletion.
- 19. An oligonucleotide primer for the PCR and the sequencing of exon 1 and/or 5'-untranslated region of the PTEN/MMAC1 gene and its homologues, characterized by the following sequences:

PTENex1-401 sense 5'-ccttctactgcctcca -3' PTENex1 -465 sense 5'- gggagggggtctgagt -3'

PTENex1 ATG sense 5'- atgacagccatcatcaaaga -3'

PTENex1 R antisense 5'- aggtcaagtctaagtcgaatc -3'

20. The oligonucleotide primer for PCR and the sequencing of exon 2 of the PTEN/MMAC1 gene and its homologues, characterized by the following sequences:

PTENex2F sense
5'- atatttatccaaacattattgctat -3'

PTENex2R antisense
5'- cttactacatcatcatattgttcc -3'

21. The oligonucleotide primer for PCR and the sequencing of exon 4, intron 4 and exon 5 of the PTEN/MMAC1 gene and its homologues, characterized by the following sequences:

Zoo43sUV sense 5'- tqtqctgagagacattatgac -3'

SPL5 sense 5'- aaatttaattgcagaggt -3'

Zoo44aRV antisense 5'- ttgtctctggtccttacttc -3'

22. The oligonucleotide primer for PCR and the sequencing of exon 5 of the PTEN/MMAC1 gene and its homologues, characterized by the following sequences:

PTEN se sense
5'- atcttgaccaatggctaagtg -3'

Zoo44aRV antisense

5'- ttgtctctggtccttacttc -3'

23. The oligonucleotide primer for PCR and the sequencing of exon 6 of the PTEN/MMAC1 gene and its homologues, characterized by the following sequences:

PTENex6F sense

5'- gga gta act att ccc agt cag ag -3'

PTENex6R antisense

5'- gca agt tcc gcc act gaa -3'

24. The oligonucleotide primer for PCR and the sequencing of exon 7 of the PTEN/MMAC1 gene and its homologues, characterized by the following sequences:

PTENex7F sense

5'- cct cag ttt gtg gtc tgc ca -3'

PTENex7R antisense

5'- c ctt ttt tag cat ctt gtt ctg ttt -3'

25. The oligonucleotide primer for PCR and the sequencing of exon 8 of the PTEN/MMAC1 gene and its homologues, characterized by the following sequences:

PTENex8F sense

5'- caa aat gtt tca ctt ttg ggt aaa -3'

PTENex8R antisense

5'- taa aat ttg gag aaa agt atc ggt t -3'

26. The oligonucleotide primer for PCR and the sequencing of exon 9 of the PTEN/MMAC1 gene and its homologues, characterized by the following sequences:

PTENex9F sense

5'- gtg aag ctg tac ttc aca aaa ac -3'

#### PTENex9tga antisense

- 5'- aaa aaa att cag act ttt gta att tg -3'
- 27. The method according to claims 1 to 17, characterized in that for the DNA amplification a mixture of oligonucleotides is used which differ at the 3' region of the oligonucleotide as regards its length by one or more nucleotides or which differ as regards their nucleotide sequence at the 3' end of the oligonucleotide at one or more positions.
- 28. The method according to claims 1 to 17 and 26, wherein the oligonucleotides

#### sense:

- 5'- cga cgt tgt aaa acg acg gcc agt tgt gct gag aga cat tat gac -3',
- 5'- cga cgt tgt aaa acg acg gcc agt tgt gct gag aga cat tat -3',
- 5'- cga cgt tgt aaa acg acg gcc agt tgt gct gag aga cat t -3',

#### antisense:

- 5'- cag gaa aca gct atg act tgt ctc tgg tcc tta ctt c -3',
- 5'- cag gaa aca gct atg act tgt ctc tgg tcc tta c -3',
- 5'- cag gaa aca gct atg act tgt ctc tgg tcc t. -3'

are used for the amplfication.

29. The method according to claims 1 to 17 and 26, wherein the oligonucleotides

#### sense:

- 5'- cga cgt tgt aaa acg acg gcc agt tgt gct gag aga cat tat gaa -3',
- 5'- cga cgt tgt aaa acg acg gcc agt tgt gct gag aga cat tat gac -3',

- 5'- cga cgt tgt aaa acg acg gcc agt tgt gct gag aga cat tat gag -3',
- 5'- cga cgt tgt aaa acg acg gcc agt tgt gct gag aga cat tat gat -3',

#### antisense:

- 5' cag gaa aca gct atg act tgt ctc tgg tcc tta ctt a -3',
- 5'- cag gaa aca gct atg act tgt ctc tgg tcc tta ctt c -3',
- 5'- cag gaa aca gct atg act tgt ctc tgg tcc tta ctt g -3',
- 5'- cag gaa aca gct atg act tgt ctc tgg tcc tta ctt t -3'

are used for the amplification.

- 30. The method according to claims 1 to 17, characterized in that DNA sequencing methods are used for the genetic analysis.
- 31. The method according to claims 1 to 17, characterized in that DNA sequencing techniques are used in the genetic analysis for the PTEN/MMAC1 and/or its pseudogenes and their homologues.
- 32. The method of distinguishing the DNA of various species, characterized in that at least one hybridization probe pair is used, the melting points of different combinations are determined and compiled for each species into a panel.
- 33. The method of distinguishing the DNA of various species, characterized in that at least one hybridization probe pair is used and at least one gene segment is amplified, differing hybridization probe pairs hybridize to different gene segments, and the melting points of the different combinations are determined and compiled for each species into a panel and/or compared with this panel for the purpose of identification.

- 34. The method of distinguishing the DNA of different species according to claim 33, characterized in that at least one hybridization probe pair is used and at least one gene segment of at least one species is amplified, differing hybridization probe pairs hybridize to different gene segments of various species, and the melting points of the different combinations are determined and compiled for each species into a panel and/or compared with this panel for the purpose of identification.
- 35. The method of distinguishing the DNA of various species according to claims 33 and 34, characterized in that at least two hybridization probes of SEQ Nos. 3 to 8 are used, the melting points of different combinations are determined and compiled for each species into a panel.
- 36. The method according to claims 33 and 34, characterized in that the species differentiation of pig DNA from various other species is made using the hybridization probe pair A1/A2 as the hybridization probe pair.
- 37. The method according to claims 33 and 34, characterized in that the hybridization probes are used in combinations C1/C2; A1/B2; A1/A2; C1/A2; B1/B2; B1/A2 for the species differentiation between various species.
- 38. LightCycler hybridization probes for exon 5, characterized by the sequences:
  - A1: 5'- tgc ata ttt gtt tca tcc ggg caa att fluorescein -3'
  - A2: 5'- LC Red 705 tta aag gca caa gat ttc tat ggg ga ph -3'
  - B1: 5'- tgc ata ttt att aca tcg ggg caa att fluorescein -3'
  - B2: 5'- LC Red 640 aag gca caa gag gcc cta gat ttc ta ph -3'

C1: 5'- tgc ata ttt gtt aca tcg ggg taa att - fluorescein -3'

C2: 5'- LC Red 640 - aag gca caa gag gcc cta gat ttc ta - ph -3'

39. LightCycler hybridization probes for exon 6, characterized by the sequences

#### PTENex6FL

5'- tca tct gga tta tag acc agt ggc act - fluorescein -3'

#### PTENex6LC 640

5'-LC Red 640-ttc aca aga tga tgt ttg aaa cta ttc caa- ph -3'

#### PTENex6F\*

5'- gtg cca ctg gtc tat aat cca gat- fluorescein -3'

### PTENex6L\* 705

5'- LC Red 705- ttc ttt aac agg tag cta taa taa tac aca ta- ph -3'

40. The LightCycler hybridization probes for exon 7, characterized by the sequences

## PTENex7F\*

5'- taa agg tga aga tat att cct cca att ca - fluorescein -3'

#### PTENex7L\*640

5'-LC Red 640- acc cac acg acg gga aga caa g - ph -3'

#### PTENex7 FL

5'-ggtaacggctgagggaactcaaagtac - fluorescein -3'

#### PTENex7 LC (705-labeled)

5'-LC Red 705- tgaacttgtcttcccgtcgtgtgg- ph -3'

41. The LightCycler hybridization probes for exon 8, characterized by the following sequences

PTENex8F\*

5'- tga caa gga ata tct agt act tac ttt aac aaa-fluorescein -3'

PPTENex8L\* 705

5'-LC Red 705 - ctt gac aaa gca aat aaa gac aaa gc- ph - 3'

PTENex8 FLU

5'- tgctatcgatttcttgatcacatagacttccatttt - fluorescein - 3'

PTENex8 LCR (640-labeled)

5'-LC Red 640- actttttctgaggtttcctctggtcctggtat - ph -3'

42. The LightCycler hybridization probes for exon 9, characterized by the following sequences

PTENex9 FL

5'-aac atc tgg tgt tac aga agt tga act gct- fluorescein -3'

PTENex9 LC 640

5'-LC-640- cct ctg gat ttg acg gct cct cta ct - ph -3'

43. Hybridization probe pair A1/A2: specific to PTEN pseudogene pig, characterized by

SEQ No. 3 Al: 5'- tgc ata ttt gtt tca tcc ggg caa att - fluorescein -3'

SEQ No. 4 A2: 5'-LC Red 705- tta aag gca caa gat ttc tat ggg ga - ph -3'

44. Hybridization probe pair B1/B2: specific to pseudogene man, characterized by

SEQ No. 5 B1: 5'- tgc ata ttt att aca tcg ggg caa att - fluorescein -3'

SEQ No. 6 B2: 5'-LC Red 640- aag gca caa gag gcc cta gat ttc ta -ph -3'

45. Hybridization probe pair C1/C2: specific to PTEN pseudogene man (C2) and homologue of pig (C1), characterized by

SEQ No. 7 - C1: 5'- tgc ata ttt gtt aca tcg ggg taa att - fluorescein -3'

SEO No. 8 - C2: corresponds to probe B2.

- 46. DNA sequences and/or fragments of homologues of the PTEN/MMAC1 gene and/or of the homologues of the PTEN/MMAC1 pseudogene, which code for proteins involved in the cell-cell adhesion and cell cycle regulation and have an indispensable function in embryogenesis of the respective species and which are compiled in the annex under "List of species sequences".
- 47. DNA sequences of homologues of the PTEN/MMAC1 gene and/or of homologues of the PTEN/MMAC1 pseudogene, which are compiled in the annex under "list of species sequences", which as compared to the PTEN/MMAC1 gene and/or the PTEN/MMAC1 pseudogene comprise genetic variants such as base substitutions and/or insertions and/or deletions and are suited for identifying corresponding species.
- 48. A kit for carrying out the method according to claims 1 to 18 and further claims, comprising:
  - a) one or more vessels comprising PCR and/or sequencing oligonucleotides binding to highly conserved genes, the oligonucleotides being optionally labeled radioactively or by means of a

dye or in another way,

b) vessels having further common reagents for DNA amplification and/or DNA analysis, in particular for DNA sequencing,

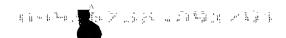
and

- c) a vessel containing a control DNA which is suited for testing the oligonucleotides and the reaction conditions.
- 49. The kit according to claim 48 for carrying out the method according to claims 1 to 18 and further claims, comprising:
  - a) one or more vessels with PCR and/or sequencing oligonucleotides according to claims 19 to 26.
- 50. Kit for identifying species for carrying out the method according to claims 1 to 18 and further claims, comprising:
  - a) a vessel having an oligonucleotide pair comprising the following sequences:
    - 5'- cga cgt tgt aaa acg acg gcc agt tgt gct gag aga cat tat gac -3' and 5'- cag gaa aca gct atg act tgt ctc tgg tcc tta ctt c -3',
  - b) two vessels with one of the following sequencing oligonucleotides each, these oligonucleotides being optionally labeled radioactively or by means of a dye or in another way:
    - 5'- cag gaa aca gct atg ac -3' and
    - 5'- cga cgt tgt aaa acg acg gcc agt -3',
  - c) a vessel containing a control DNA, which is suited for testing the oligonucleotides and the reaction conditions.
- 51. The kit (Light Cycler Kit) for carrying out the method according to claims 32 to 37 and further claims, comprising
  - a) one or more vessels containing PCR primers and hybridization probes, which bind to highly conserved genes, the hybridization probes being

- optionally labeled by means of a dye,
- b) vessels containing further common reagents for DNA amplification and/or DNA analysis, in particular for the Light Cycler Analyses,

and

- c) a vessel containing a control DNA which is suited for testing the oligonucleotides and the reaction conditions.
- 52. The kit (Light Cycler Kit) for carrying out the method according to claims 32 to 37 and further claims, comprising:
  - a) one or more vessels with PCR primers and hybridization probes according to claims 38 to 42.



(12) NACH DEM VERTRAG ÜBER DIE INTERNATIONALE ZUSAMMENARBEIT AUF DEM GEBIET DES PATENTWESENS (PCT) VERÖFFENTLICHTE INTERNATIONALE ANMELDUNG

(19) Weltorganisation für geistiges Eigentum Internationales Büro



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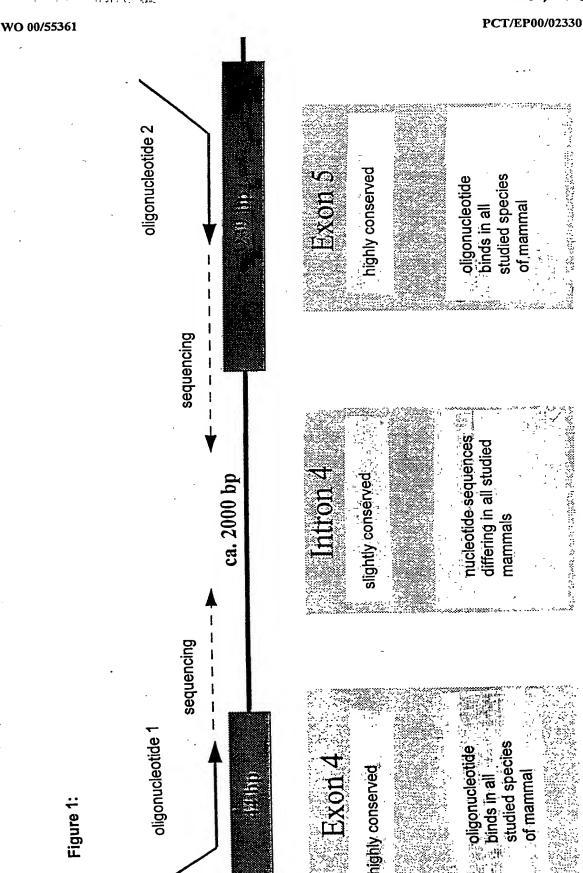
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#### Veröffentlicht:

mit internationalem Recherchenbericht

[Fortsetzung auf der nachsten Seite]

- (54) Title: METHOD FOR IDENTIFYING ORGANISMS BY MEANS OF COMPARATIVE GENETIC ANALYSIS AND PRIMERS AND HYBRIDISATION PROBES FOR CARRYING OUT THIS METHOD
- (54) Bezeichnung: VERFAHREN ZUR IDENTIFIZIERUNG VON ORGANISMEN DURCH VERGLEICHENDE GENETI-SCHE ANALYSE SOWIE PRIMER UND HYBRIDISATIONSSONDEN ZUR DURCHFÜHRUNG DES VERFAHRENS
- (57) Abstract: The invention comprises methods, primers and hybridisation probes for identifying organisms by means of comparative genetic analysis and is characterised in that coding and non-coding areas and/or functionally significant areas of highly conserved genes, pseudogenes or homologues are amplified using the PCR and then genotyped and analysed. The comparison of coding and non-coding areas of highly conserved genes, pseudogenes or homologues ensures that a single oligonucleotide pair bonds to DNA sequences that are highly conserved between different species, hereby allowing the amplification of a gene segment that is identical for all of the species. The oligonucleotides include one or more gene areas with the greatest possible sequence differences between different species. The determination of the gene sequence of this highly polymorphous area in a subsequent reaction step enables the gene sequence to be allocated to a specific species. In particularly preferred variants of embodiments of the invention, oligonucleotide pairs are found that make it possible to amplify the highly conserved tumour suppresser gene PTEN/MMAC1, its pseudogene and their homologues.
- (57) Zusammenfassung: Die Erfindung umfaßt Verfahren, Primer und Hybridisationssonden zur Identifizierung von Organismen durch vergleichende genetische Analyse, dadurch gekennzeichnet, daß man kodierende, nichtkodierende Bereiche und/oder funktionell bedeutende Bereiche von hochkonservierten Genen, Pseudogenen oder Homologen mit Hilfe der PCR amplifiziert und nachfolgend genotypisiert und analysiert. Der Vergleich von kodierenden und nicht kodierenden Bereichen von hochkonservierten Genen, Pseudogenen oder Homologen gewährleistet, daß ein einziges Oligonukleotidpaar an DNA-Sequenzen, die zwischen verschiedenen Spezies hochgradig konserviert sind, bindet und damit die Amplifikation eines für alle Spezies identischen Genabschnitts erlaubt. Die Oligonukleotide schließen einen oder mehrere Genbereiche ein, die zwischen verschiedenen Spezies möglichst große Sequenzunterschiede aufweisen. Die Bestimmung der Gensequenz dieses hochgradig polymorphen Bereichs in einem darauf folgenden Reaktionsschritt erlaubt es, die Gensequenz einer spezifischen Spezies zuzuordnen. In besonders bevorzugten Ausführungsvarianten der Erfindung wurden Oligonukleotidpaare gefunden, die die Amplifikation des hochkonservierten Tumorsuppressorgens PTEN/MMAC1, seines Pseudogens und ihrer Homologen ermöglicht.



1/8 **ERSATZBLATT (REGEL 26)** 

man African elephant	TCT	5H.ΩΩ	20 	30 CA	30ACTCTA.CCG-	%	G	0
man African elephant	T		% G.ACA. A.GTG.		9	120	GG	. A-
man African elephant	150	150	. G T	170	180 A	061	260 	210
man African elephant	220 	220 A	230	240	250	260	270	580
man African elephant	290	290	300	310	320 	330 AT	340 AG	350

pig pig liver beef salami		D D H	2	20		6   	E H H D	8 8 H H D	
pig pig liver beef salami		HHU	· · · · · · · · · · · · · · · · · · ·	06	001	110	120	130	140
pig pig liver beef salami	-	Ü Ü Ü		91	170 T T		180 G.TCG.TGTG	500	210
pig pig liver beef salami	•		220 	230		,			

Figure 4:	ure 4: Positions of the hybridization probes in exon 5 of the PTEN gene,
•	nseudogene and their homologues

220 SATTICTA TOGGGAAGT		TO A A OCOUNTY A POTENTY AV			- TTTCTATGGGGA AGT
B1 TATTACATCGGGGAAATT TITA AAGGCACAGAGGGCCCTAGATTTCTA TGGGGAAGT				A2	TTGTTTCATCCGGGCAAATT TTAAAGGCACAAGA
150 ATVATO TOOATATTTATTACATOGGGGAAAT		TO	ATTIC ICCATATION ACATOGOGGIAAAN	A1	ATITIC TCCATATITGTTTCATCCGGGCAAAT
150	Č	į	<		ΛT
•	pseudogene		pig gene		pig pseudogene

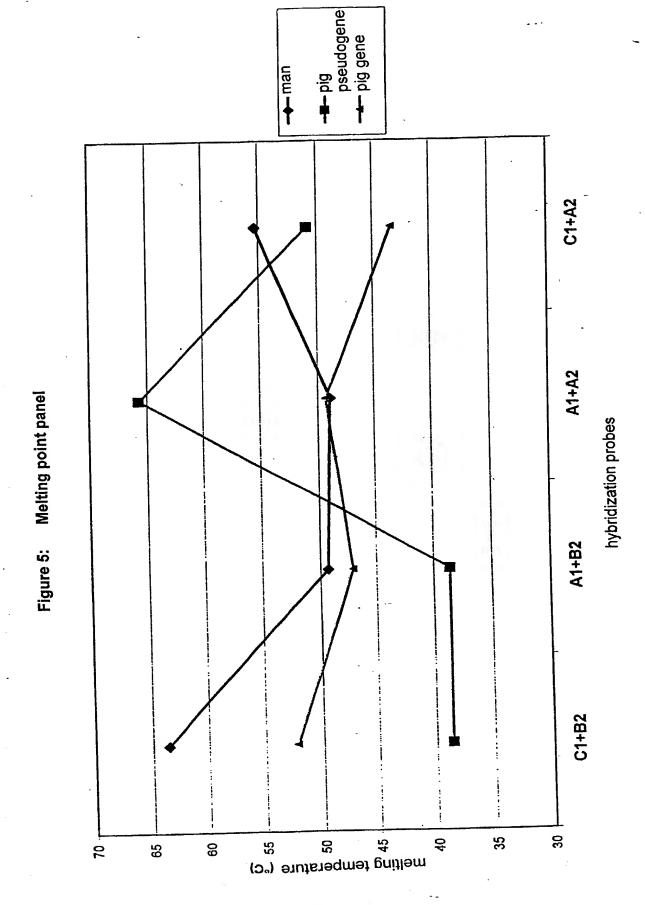
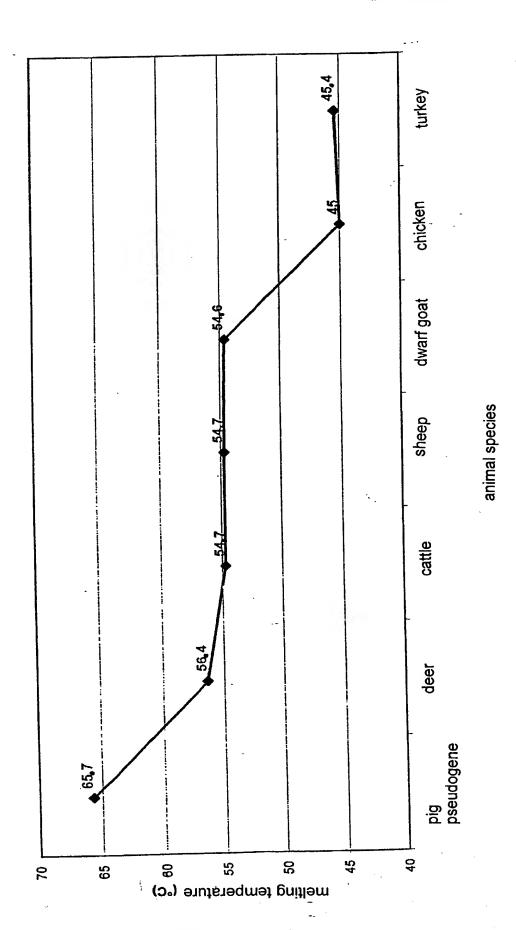


Figure 6: Melting point with probe A1 + A 2



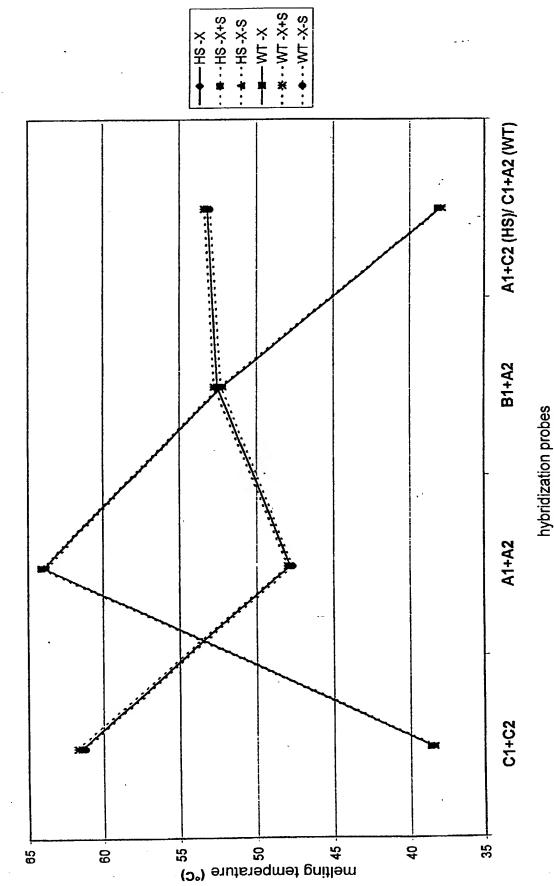
- 83 - 21 - 34

hybridization probes

pig pseudogene goitred gazelle guinea pig esnow deer \* trout duck ay duail gob B1+A2 B1+B2 C1+A2 Figure 7: Melting point panel A1+A2 A1+B2 C1+B2 6 35 39 45 55 22 65 9 melting temperature (°C)

Figure 8: Standard deviations of select probes





10 Res 2007 1 9 FEB 2007 Attorney's Docket No. 012627-025

#### IN THE UNITED STATES PATENT AND TRADEMARK OFFICE

In re Patent Application of	)
Hans Konrad SCHACKERT et al.	) Group Art Unit: Unassigned
Serial No.: 09/936,738	) Examiner: Unassigned
Corresponding to PCT/EP00/02330	)
International Filing Date: 3 March 2000	) ATTENTION: BOX SEQUENCE
For: METHOD FOR IDENTIFYING	)
ORGANISMS BY MEANS OF	)
COMPARATIVE GENETIC	)
ANALYSIS AND PRIMERS AND	)
HYBRIDISATION PROBES FOR	)
CARRYING OUT THIS METHOD	)

# DECLARATION PURSUANT TO 37 C.F.R. §§ 1.821-1.825

Assistant Commissioner for Patents Washington, D.C. 20231

Sir:

- I, Deborah H. Yellin, declare as follows:
- That the content of the paper and computer readable copies of the Sequence
  Listing, submitted in accordance with 37 C.F.R. § 1.821(c) and (e),
  respectively, are the same in compliance with § 1.821(f).
- 2. That the submission, filed in accordance with 37 C.F.R. § 1.821(g)[or (h)], herein does not include new matter [or go beyond the disclosure in the international application].

N9936733.C91.701.

Serial No.: 09/936,738

I hereby declare that all statements made herein of my own knowledge are true and that all statements were made on information and belief and are believed to be true; and further that these statements were made with the knowledge that willful false statements and the like so made are punishable by fine or imprisonment, or both, under Section 1001 of Title 18 of the United States Code and that such willful false statements may jeopardize the validity of the application or any patent issued thereon.

February 22, 2002

Date

Debuyella Deborah H. Yellin

Registration No. 30,427

COMBINED DECLARATION FOR (Includes Reference to Provision	PATENT APPLICATION AND Fal and PCT International Applic	POWER OF ATTORNEY ations)	Attorney's Docket No. 012627-025		
As a below named inventor, I he My residence, post office addres I believe I am the original, first a (if plural names are listed below) entitled:	s and citizenship are as stated b	ame is listed below) or an oris	zinal, first and joint inventor		
METHOD FOR IDENTIFYING	G ORGANISMS BY MEANS O	F COMPARATIVE GENETI	C ANALYSIS AND		
PRIMERS AND HYBRIDIZAT	ION PROBES FOR CARRYIN	IG OUT THIS METHOD			
the specification of which	ch (check only one item below)	:			
is attached hereto					
Number	ed States application				
on and was amended					
on		(if applicable).			
Number <u>PCT/F</u> on <u>16 March 20</u> and was amended	000	(if applicable).			
I hereby state that I have review amended by any amendment ref	ed and understand the contents erred to above.	of the above-identified-specifi			
I acknowledge the duty to disclose to the Office all information known to me to be material to patentability as defined in Title 37, Code of Federal Regulations, §1.56.					
I hereby claim foreign priority benefits under Title 35, United States Code, §119 (a)-(e) of any foreign application(s) for patent or inventor's certificate or of any PCT international application(s) designating at least one country other than the United States of America listed below and have also identified below any foreign application(s) for patent or inventor's certificate or any PCT international application(s) designating at least one country other than the United States of America filed by me on the same subject matter having a filing date before that of the application(s) of which priority is claimed:					
PRIOR FOREIGN/PCT APPLI	CATION(S) AND ANY PRIOF				
COUNTRY (if PCT, indicate "PCT")	APPLICATION NUMBER	DATE OF FILING (day, month, year)	PRIORITY CLAIMED UNDER 35 U.S.C. §119		
Germany	199 11 656.3	16-03-99	_X_Yes No		
Germany	199 64 112.9	31-12-99	_X_Yes _ No		
			_ Yes _ No		
			_ Yes No		
			_ Yes _ No		
I hereby claim the benefit under below.	r Title 35, United States Code §	§ 119(e) of any United States p	provisional application(s) listed		
(Application Nu	umber)	(Filing Date)			
(Application Nu	ımber)	(Filing Date)			

COMBINED DECLARATION FOR PATENT APPLICATION AND POWER OF ATTORNEY (CONT'D)	Attorney's Docket No.	
(Includes Reference to Provisional and PCT International Applications)	012627-025	

I hereby claim the benefit under Title 35, United States Code, §120 of any United States applications(s) or PCT international application(s) designating the United States of America that is/are listed below and, insofar as the subject matter of each of the claims of this application is not disclosed in that/those prior application(s) in the manner provided by the first paragraph of Title 35, United States Code, §112, I acknowledge the duty to disclose to the Office all information known to me to be material to the patentability as defined in Title 37, Code of Federal Regulations §1.56, which became available between the filing date of the prior application(s) and the national or PCT international filing date of this application:

PRIOR U.S. APPLICATIONS OR PCT INTERNATIONAL APPLICATIONS DESIGNATING THE U.S. FOR BENEFIT UNDER 35 U.S.C. §120:

U.S. APPLICATIONS			STATUS (check one)		
U.S. APPLICATION N	UMBER	U.S. FILING DATE	PATENTED	PENDING	ABANDONED
PCT	APPLICATIONS DESIGNATIN	G THE U.S.			
PCT APPLICATION NO.	PCT FILING DATE	U.S. APPLICATION NUMBERS ASSIGNED (if any)			
					1

I hereby appoint the following attorneys and agent(s) to prosecute said application and to transact all business in the Patent and Trademark Office connected therewith and to file, prosecute and to transact all business in connection with international applications directed to said invention:

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Norman H. Stepno	22,716	William C. Rowland	30,888	Allen R. Baum
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Samuel C. Miller, III	27,360	Peter K. Skiff	31,917	Mary Ann Dillahunty
Robert G. Mukai	28.531	Richard J. McGrath	29,195	6 1
George A. Hovanec, Jr.	28,223	Matthew L. Schneider	32,814	
James A. LaBarre	28,632	Michael G. Savage	32,596	21839
E. Joseph Gess	28,510	Gerald F. Swiss	30.113	
R Danny Huntington	27.903	Charles F. Wieland III	33,096	

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I hereby declare that all statements made herein of my own knowledge are true and that all statements made on information and belief are believed to be true; and further that these statements were made with the knowledge that willful false statements and the like so made are punishable by fine or imprisonment, or both, under Section 1001 of Title 18 of the United States Code and that such willful false statements may jeopardize the validity of the application or any patent issued thereon.

Page 2 of 3

33,815 34,040 31,979 36,341 36,086

32,747 36,075 32,236 34,456 34,576

Attorney's Docket No. COMBINED DECLARATION FOR PATENT APPLICATION AND POWER OF ATTORNEY (CONT'D) (Includes Reference to Provisional and PCT International Applications) 012627-025 FULL NAME OF SOLE OR FIRST INVENTOR 5-01 Hans Konrad Schackert RESIDENCE CITIZENSHIP Zittauer Straße 17, D-01099 Dresden, Germany DEX German POST OFFICE ADDRESS Zittauer Straße 17, D-01099 Dresden, Germany FULL NAME OF SECOND JOINT INVENTOR, IF ANY SIGNATURE DATE 18.12.01 Matthias Hahn RESIDENCE CITIZENSHIP Jahnstraße 14, D-68526 Ladenburg, Germany German POST OFFICE ADDRESS Jahnstraße 14, D-68526 Ladenburg, Germany SIGNATURE HOUL FULL NAME OF THIRD JOINT INVENTOR, IF ANY DATE 1201 Olga Niki Koufaki RESIDENCE CITIZENSHIP Senefelder Straße 2/Zi. 620, D-01307 Dresden, Germany Greek POST OFFICE ADDRESS Senefelder Straße 2/Zi. 620, D-01307 Dresden, Germany FULL NAME OF FOURTH JOINT INVENTOR, IF ANY SIGNATURE DATE 5-01 Heike Görgens RESIDENCE CITIZENSHIP Weinbergstraße 50, D-01129 <u>Dresden</u>, Germany German POST OFFICE ADDRESS Weinbergstraße 50, D-01129 Dresden, Germany FULL NAME OF FIFTH JOINT INVENTOR, IF ANY SIGNATURE DATE RESIDENCE CITIZENSHIP POST OFFICE ADDRESS FULL NAME OF SIXTH JOINT INVENTOR, IF ANY SIGNATURE DATE RESIDENCE CITIZENSHIP POST OFFICE ADDRESS FULL NAME OF SEVENTH JOINT INVENTOR, IF ANY **SIGNATURE** DATE RESIDENCE CITIZENSHIP POST OFFICE ADDRESS FULL NAME OF EIGHTH JOINT INVENTOR, IF ANY SIGNATURE DATE RESIDENCE CITIZENSHIP POST OFFICE ADDRESS

## SEQUENCE LISTING

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<400> 77 agccatcatc cttagac	aaagagatcg	ttagcagaaa	caaaaggaga	tatcaagagg	atggattcga	60 67
<210> 78 <211> 69 <212> DNA <213> Rabbi	ít					
<400> 78 acagccatca gacttagac	tcaaagagat	cgttagcaga	aacaaaagga	gatatcaaga	ggatggattc	60 69
<210> 79 <211> 65 <212> DNA <213> Hare						
<400> 79 cagccatcat actta	caaagagatc	gttagcagaa	acaaaaggag	atatcaagag	gatggattcg	60 65
<210> 80 <211> 59 <212> DNA <213> Antel	Lope					
<400> 80 ccatcatcaa	agagatcgtt	agcagaaaca	aaaggagata	tcaagaggat	ggattcgac	59
<210> 81						

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<211> 65
<212> DNA
<213> Kangaroo
<400> 81
gccatcatca aagagatcgt gagcagaaac aaaaggagat accaagagga tqqattcgac 60
ttaga
<210> 82
<211> 25
<212> DNA
<213> Artificial Sequence
<220>
<223> PTENex2F sense
<400> 82
atatttatcc aaacattatt gctat
                                                                    25
<210> 83
<211> 25
<212> DNA
<213> Artificial Sequence
<220>
<223> PTENex2R antisense
<400> 83
                                                                    25
cttactacat catcaatatt gttcc
<210> 84
<211> 69
<212> DNA
<213> Man
<400> 84
tocaaacatt attgctatgg gatttcctgc agaaagactt gaaggcgtat acaggaacaa 60
                                                                    69
tattgatga
<210> 85
<211> 69
<212> DNA
<213> Chimpanzee
<220>
<221> misc feature
<222> (1) ... (69)
<223> n = A,T,C or G
<400> 85
aaacattatt gctatgggat ttcctgcaga aagacttgaa ggcgtatana ggaacaatat 60
tgatgatgt
<210> 86
<211> 70
<212> DNA
<213> Domestic pig
```

```
<400> 86
ccaaacatta ttgctatggg gtttcctgca gaaagacttg aaggcgtata caggaacaat 60
attgatgatg
<210> 87
<211> 71
<212> DNA
<213> Wild boar
<400> 87
aaacattatt gctatggggt ttcctgcaga aagacttgaa ggcgtataca ggaacaatat 60
tgatgatgta g
<210> 88
<211> 63
<212> DNA
<213> Cattle
<400> 88
cattattgct atgggctttc ctgcagaaag acttgaaggt gtatacagga acaatattga 60
<210> 89
<211> 62
<212> DNA
<213> Sheep
<400> 89
ttattgctat ggggtttcct gcagaaagac ttgaaggcgt gtacaggaac aatattgatg 60
<210> 90
<211> 58
<212> DNA
<213> Goat
<400> 90
ttattgctat ggggtttcct gcagaaagac ttgaaggcgt gtacaggaac aatattga 58
<210> 91
<211> 64
<212> DNA
<213> Red buffalo
<220>
<221> misc_feature
<222> (1) ... (64)
<223> n = A,T,C or G
<400> 91
cattattgct atggggtttc ctgcagaaag acttgaaggc gtatnnagga acaatattga 60
tgat
                                                                    64
<210> 92
<211> 68
<212> DNA
<213> Deer
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```
<400> 92
tttatccaaa cattattgct atggggtttc ctgcagaaag acttgaaggc gtatacagga 60
acaatatt
<210> 93
<211> 58
<212> DNA
<213> Roe deer
<220>
<221> misc_feature
<222> (1) ... (58)
\langle 223 \rangle n = A,T,C or G
<400> 93
ttattgctat ggggtttcct gcagaaagac ttgaaggcgt atannggaac aatattga
<210> 94
<211> 65
<212> DNA
<213> Goitred gazelle
<400> 94
ccaaacatta ttgctatggg gtttcctgca gaaagacttg aaggcgtata caggaacaat 60
attga
<210> 95
<211> 64
<212> DNA
<213> Horse
<400> 95
attattgcta tggggtttcc tgcagaaaga cttgaaggcg tatacaggaa caatattgat 60
<210> 96
<211> 67
<212> DNA
<213> Dog
<220>
<221> misc_feature
<222> (1) ... (67)
<223> n = A, T, C or G
<400> 96
ttccaaacat tattgctatn gggtttcctg cagaaagact tgaaggcgta tacnggaaca 60
atattga
<210> 97
<211> 65
<212> DNA
<213> Sun bear
<220>
<221> misc feature
<222> (1) ... (65)
```

```
<223> n = A, T, C or G
<400> 97
tccaaacatt attgctatng ggtttcctgc agaaagactt gaaggcgtat acaggaacaa 60
tattg
<210> 98
<211> 62
<212> DNA
<213> Rabbit
<400> 98
gctatgggat ttcctgcaga aagacttgaa ggcgtataca ggaacaatat tgatgatgta 60
gt
                                                                    62
<210> 99
<211> 59
<212> DNA
<213> Hare
<400> 99
acattattgc tatgggattt cctgcagaaa gacttgaagg cgtatacagg aacaatatt 59
<210> 100
<211> 48
<212> DNA
<213> Antelope
ttgctatggg gtttcctgca gaaagacttg aaggcgtata caggaaca
                                                                    48
<210> 101
<211> 77
<212> DNA
<213> Turkey
<400> 101
tttatccaaa cattattgct atgggttttc ctgcggagag gcttgaagga gtataccgga 60
acaatattga tgatgta
<210> 102
<211> 73
<212> DNA
<213> Chicken
<400> 102
atttatccaa acattattgc tatgggtttt cctgcggaga ggcttgaagg agtataccgg 60
aacaatattg atg
                                                                    73
<210> 103
<211> 61
<212> DNA
<213> Duck
<400> 103
ttattgctat gggttttcct gcagagaggc ttgaaggagt gtaccggaac aatattgatg 60
                                                                    61
```

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<210> 104
<211> 62
<212> DNA
<213> Quail
<400> 104
cattattgct átgggttttc ctgcggagag gcttgaagga gtataccgga acaatattga 60
tg
<210> 105
<211> 73
<212> DNA
<213> Goose
<400> 105
tttatccaaa cattattgct atgggttttc ctgcagagag gcttgaagga gtgtaccgga 60
acaatattga tga
<210> 106
<211> 66
<212> DNA
<213> Ostrich
<400> 106
ccaaacatta ttgctatggg ttttccggcg gagaggcttg aaggagtgta ccggaacaat 60
attgat
                                                                    66
<210> 107
<211> 59
<212> DNA
<213> Pigeon
<400> 107
cattattgct atgggttttc ctgcggagag gcttgaagga gtataccgga acaatattg 59
<210> 108
<211> 60
<212> DNA
<213> Varan
<400> 108
cattattgct atgggttttc ctgcggagag gcttgaagga gtataccgga acaatattga 60
<210> 109
<211> 21
<212> DNA
<213> Artificial Sequence
<220>
<223> Zoo43sUV
<400> 109
tgtgctgaga gacattatga c
                                                                    21
<210> 110
<211> 20
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<212> DNA
<213> Artificial Sequence
<220>
<223> Zoo44aRV
<400> 110
ttgtctctgg tccttacttc
                                                                     20
<210> 111
<211> 654
<212> DNA
<213> Man
<400> 111
ttatgacacc gccaaattta attgcagagt atgaatgtac tgtactatgt tgtataactt 60
aaacccgata gactgtatet tactgtcata acaataatga gtcatccaga ttatcgagtg 120
agatacatat ttaagaatta totttaaaaaa tttoaaaaat tttaatttta otgttgtgtt 180
ttaggaaaaa gtattgcata aagctattaa tattgtcagg aagactaaag tgcagcatag 240
actaagaatt aggaaaattc ctagactaaa aatagtataa ggagagggtt tacctactat 300
ttgaggcagt tggtctaata gtaagcaatc acagggagaa agcagaacta cttaactctt 360
ctgtgttgag gaatgacata aaaggtagga aaggatataa caaatgttga taagaggagt 420
ctgatggatg agaggaggga actgctttaa atgagtttct acttcagaca taagttaatt 480
ctcagagccc acaaaaactt tcacttttat ttgtgaaata caactcagtt ctcatggctt 540
aacactttaa accatgagaa aactgaagag ttgagagctt ggcagatgct gctgtgatag 600
tcaaaagaaa gtgggtgcat gagctactat tgatgtattt gccatggtcc ctcc
<210> 112
<211> 582
<212> DNA
<213> Dog
<400> 112
atgtaataaa tatgcacaaa tcattacacc agttcgtccc tttccagctt tacagtgaat 60
tgctgcaaca tgattgtcat cttcacttag ccattggtca agatcttcac aaaagggttt 120
gataagttct agctgtggtg gattatggtc ttcaaaagga tactgtgcaa ctgtggtaaa 180
aagataacct cagaattaga aaaaagtctt tootgaactg tttattaaaa gtaggttaac 240
tttagaaaca ttgcatgtaa gcttaacaga tgtttaaaag aaaaacggaa ctccagagaa 300
aaataatttg ctgtctgata attttccaat ttttgaatag aaaatagtct ctcattaatt 360
cttaaaccta ccactadgag agagaggcta agcattattt tcccccactt taatgaaaga 420
ggaaactttg caatggagag ggagcacacg tcaacatatc agagggaaga ggcaaactca 480 aaatgaaatg gcacacaggt ttcctgtcag ggctctcaat gcattttctg acaaaaggag 540
tcataatatt tataatacta cgtcatccaa aatatatatt cc
<210> 113
<211> 376
<212> DNA
<213> Cattle
<220>
<221> misc feature
<222> (1)...(376)
<223> n = A,T,C or G
<400> 113
taggtacaca tattgtgtta gataacttga agccaacagt ctaaatttta ctgtcatacc 60
aataatgaat aatctcaagt attaagtgat atatttatct taaagatggt ctgagaaaat 120
```

```
ttgaaattaa ttttgctgtt gtgtttttgg aaataagtat catgtaaatg aggaagacta 180
aattgaatta actgaaaact aggagaaatt tatagactaa cagaataaat agaqqqttat 240
atctgtgatt tgaggcattt ggcatgatag taagagatta caggggagaa aggagaatgg 300
cttaattctg taatggaaca tgacctgtac agtgggaaaa ggggtataat gaantatgga 360
tnaaaaggag cctgaa
<210> 114
<211> 673
<212> DNA
<213> Mouse
<400> 114
ttatgacacc gccaaattta actgcagagg tatgtataaa cataaccaca gcatactgta 60
taactaaaga ccaatagact tgtcttttac tgcctggtga taattatcaa gattagtgag 120
ataaaaaatct taagaatggc ctttgacaat taaaaaaagt gtatttaatg ttagagttgt 180
tetttaagae etatetattg teaggaaaae taaateacag aataettgga gaggteecaa 240
gactaaacta ggattggagg tgcttattga cggtgtggga cagctagcgc tgctggaaac 300
aatcacaaga agagagcaga accattttaa cttttctaca tcgaagaatg gcataaagtt 360
aggaaaagat gtagcatcgg tctgtctgtc tgtctgtctg cctgtctgtc ttctcagaat 420
catgaagcac taaggagtaa gtaagaacag tttctggggg accgacagac ctaggctact 480
gctcattagg aaacatgcca tggttgaagg tcacttagct ttaaatgtac attttaacag 540
actottgaat gttottgtgt gccactgggg gaaatgaggt cgggagcaca gttagacaga 600
tggttaagta aaagctggcc tgcagcctct tggtgaatgt agtttgccat tgtttaccac 660
agagetttee tgt
<210> 115
<211> 411
<212> DNA
<213> Horse
<400> 115
aatgtacagt attttgttat ataactgaaa accagtagac taagtcttac tgtcacagca 60
gtaatgaata ctcttgatta ttaagtgaga taaatattta tcttaaaaag ataatcttag 120
aaaatttgaa aaataaattt aactttgctg ttgtatttta gaaaacaagt atcatataaa 180
ccaactggta gtattaggaa gactaaattg aagaatagac taagaattag gatgtaatag 240
taagagattg catggagaaa gcagaacgac ttaactctgg caaggagcgt gacctaaaag 300
gtggaaaagg gtataacaga tgtggataca aggagcctga acagatgaga gcagggaact 360
gcttcaaatg agttcttttc caagtatagt aaattgtttc tcagagccca c
<210> 116
<211> 566
<212> DNA
<213> Sheep
<220>
<221> misc feature
<222> (1) ... (566)
<223> n = A, T, C or G
<400> 116
aaaaatttqc nnnnqatqta acaaatatqc acaaatcatt acaccagttc qtccctttcc 60
agetttacag tgaattgetg caacatgatt gteatettea ettageeatt ggteaagate 120
ttcacaaaag ggtttgataa gttctaactg ttggtggatt atggtcttca aagggatact 180
gtgcaactgt gataaaaaga taaccgcaga tatatgaaaa taatctcact tgaattgctt 240
attacaagta ggctaacttt agaaatgttg catacaaata gtttaaaaaat gtctgaacta 300
tagaggaaaa gaatttattg tetgataatt ttetaatttt egaacagaaa ataatetete 360
attaactcaa atttatccat tegacaggta agacaagtat tattteetca etetatgatg 420
```

```
gaggcaatgg aggagcaaca tatcagaggt cacaacataa cggaggaaga ggcaaactca 480
gaatgaaacg togcacgago otottagcag ggototcaat acgttoctag caaaagggac 540
tggtaacatc tataatatcg cattat
                                                                   566
<210> 117
<211> 497
<212> DNA
<213> Turkey
<400> 117
aagctgcatt ttgccaggtg taaggaactg acagagacaa ccaaqaccaa agcatttcag 60
gctgaattcc cctckttcct cccacctcct ctgaacaaat ggaggttctg acagagtgga 120
gagattaatt cagaatatgt gtgcacagta cacctggcag accccacaaa gcttggctca 180
aagaacaaag atgaaacaaa ggcatgaata gagcagtaga aggatttaca aaaggacaaa 240
agatgggcag ccatttaaag gtgacagtaa tttcttaagt aaatgtcaaa actcttcaaa 300
gaagcaaggg ggataatatt catgaatact taaggetgaa acgtgaacat gttgatttgc 360
cattitggaag gitatgitte ettettatet eetetetgat agetteaata atgggeacta 420
aaatteqtte etgaaaaaat qeaaagaaat cacteagtgt etgaggaegt gttgatttea 480
catgtattga aatcagt
<210> 118
<211> 365
<212> DNA
<213> Trout
<220>
<221> misc feature
<222> (1)...(365)
<223> n = A, T, C or G
cattatgacn nnnnnnnatt caattgcaga ggattagata ttacatcaga gtgaaaccat 60
tatcactgtc tttcaggcag tcagtgaatg aatcaatctt tcactaaaaa cccacgtgtg 120
acgctaacta actgagcccg gtctctgtct gtctctctcc agttgcacaa tatccgtttg 180
aggatcacaa tccgccccag ctggagctga tcaaaccgtt ctgcgaagat cttggccttt 240
ggttaagtga agacgacaat catgtggcgg cgattcactk taaarctgga aaggacgtac 300
gggtgtcatg atctgtgctt acctgttaca ccggggcaag ttcctcaaag cacaagaagc 360
tctcq
<210> 119
<211> 656
<212> DNA
<213> Roe deer
<400> 119
qtataqqtac acttactatg ttagataact tgaggccaac agtctaaatt ttactatcat 60
accagtaatg aataatctca agtattaagt gatacagtca tettaaagat gatettagaa 120
aatttgaaat taattttgct gttgtgtttt tggaaacaag tgtcatgtaa atgagggaga 180
ctaaactgaa ttaactgaaa actaggagaa atttatagac tgacagaata aagaaagggt 240
tatatctgtg atttgaggca tttggcgtaa tagtaagaga ttacagggag aaaggagaat 300
gatttaattc tataatggaa catgacctgc acagtggaaa aagggtataa tgaaatataa 360
awaaaaggag cctgatagat gagagcaaga actgctttaa gtgaattttt ctccaggtat 420
agtatatttt atctcagagt ccacaaatac tttcatttgt ttttgtggaa ctcttagaac 480
gacgagagac caggaacatt gagaagctaa tatatttgcc attgttcctt cctaaatatt 540
tagcacagge tttcaaacag ttggtttaag aattcagaag tgctaataac tgagagcaag 600
ggtagattta ttactaagaa tgtttcattt ttggtggatt ttgctatttc tggtca
```

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<210> 120
<211> 405
<212> DNA
<213> Deer
<220>
<221> misc feature
<222> (1)...(405)
<223> n = A, T, C or G
<400> 120
gtataggtac actttnnaag ccaacagtct aaattttact gtcataccaa taatgaataa 60
totcaagtat taagtgatat atttatotta aagatgatot tagaaaattt gaaaotaatt 120
ttgctgttgt gtttttggaa acaagtgtca tgtaaatgag ggagaccata actgaattaa 180
ctgaaaactg ggaaaaattt atagactaac agaataaaga aagggttata tctgtggttt 240
gaggcgtttg acgtaatagt aagagattac agggagaaag gagaatgact taattctata 300
atggaacacg acctgcacag tggaaaaagg gtataatkaa atgtagataa aggagcctga 360
tagttgagag caagaactgc tttaagtgag tttttctcca ggtgt
<210> 121
<211> 522
<212> DNA
<213> Chimpanzee
<220>
<221> misc feature
<222> (1)...(522)
<223> n = A, T, C or G
<400> 121
cattatgacn nnnnnnnnn nnattgcaga ggtaggtatg aatgtactgt actatgttgt 60
ataacttaaa cocgatagac tgtatettac tgtcataaca ataatgagte atctagatta 120
 togagtgaga tacatattta tottaagaat tatotttaaa aatttoaaaa attttaattt 180
 tactcttgtg ttttaggaaa aaagtattgc ataaagctat taatattgtc aggaagacta 240
 aagtgcagca tagactaaga atgaggaaaa ttcctagact nnaatagtat aaggagaggg 300
 tttacctact atttgaggca gttggtctaa tagtaagcaa tcacagggag aaagcagaac 360
 tacttaactc ttctgtgttg aggaatgaca taaaaggtag gaaggatata acaaatgttg 420
 ataagaggag totgatggat gagaggaggg aactgettta aatgagttet actteagaca 480
 tadgttaatt ctcagagccc acaaaacttt cactittatt tg
 <210> 122
 <211> 666
 <212> DNA
 <213> Gorilla
 <220>
 <221> misc feature
 <222> (1) ... (666)
 \langle 223 \rangle n = A,T,C or G
 <400> 122
 cattatgacn nnnnnnatt taattgcaga ggtaggtatg aatgtdctgt actatgttgt 60
 ataacttaaa cccgatagac tgtatcttac tgtcataaca ataatgagtc atctagatta 120
 tcgagtgaga tacatatita tcttaagaat tatctttaaa aatttcaaaa attttaattt 180
 tactettgtg ttttaggaaa aaagtattge ataaagetat taatattgte aggaagaeta 240
 aagtgcagca tagactaaga atgaggaaaa ttcctagact nnnaatagta taaggagagg 300
 gtttacctac tatttgaggc agttggtcta atagtaagca atcacaggga gaaagcagaa 360
```

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ctacttaact cttctgtgtt gaggaatgac ataaaaggta ggraaggata taacaaatgt 420
tgataaqaqq rgtctqatgg atgagaggag ggaactgctt taaatgagtt ctacttcaga 480
cataagttaa tteteagage ceacaaaaac ttteactttt atttgtgaaa tgeaacteag 540
ttctcatqqc ttaacacttt aamccatqag agactgaaga gttgagaagc ttggcagatg 600
ctgctgtgat agtcaaaaag aaagtgggtg ccatgagcta ctattgatgt atttgccatt 660
<210> 123
<211> 741
<212> DNA
<213> Orang-utan
<220>
<221> misc feature
<222> (1)...(741)
<223> n = A, T, C or G
<400> 123
cattatgacn nnnnnaaatt taattgcaga ggtaggtacg aatgtactgt gctatgttgt 60
ataacttaaa cacaatagac tgtatettac tgtcataaca ataatgactc atctagatta 120
ttgagtgaga tacatattta tcttaagawt tatcttaaaa aatttcagaa aatttaattt 180
tactgttgtg ttttaggaaa aacgtattgc ataaagctat taatattgtc aggaaaagtg 240
cagagtagac taagaattag gaaaatteet agactaaaan nnnataagga gagggtttac 300
ctactgtttg aggcagttgg tctaatagta agcgattata gggagaaagc agaactactt 360
aactcttctg tgttgaggaa tgacatgaaa ggtaggaaag gatataacaa atgttgataa 420
gaggagcctg atggatgaga ggagggaact gctttaaatg agttctactt cagacataag 480
ttaattctca gagcccacaa aaactttcac tttcatttgt gaaatacaac tcagttctca 540
cggcttaaca ctttaaacca tgagagaact gaagagttga gaagcttggc agatgcttct 600
gtgatagtca aaaagaaagt gggtgccatg agctactatt gatgtatttg ccattgatcc 660
cycctgaaaa totagaatgg actttcagac aaatggtttg aaaatcctaa atcactaatg 720
attgggattt agtatagatt c
<210> 124
<211> 608
<212> DNA
<213> Orang-utan
<220>
<221> misc feature
<222> (1) ... (608)
<223> n = A, T, C or G
<400> 124
cattatgacn nnnncaaatt taattgcaga ggtaggtacg aatgtactgt gctatgttgt 60
ataacttaaa cacaatagac tgtatcttac tgtcataaca ataatgactc atctagatta 120
ttgagtgaga tacatattta tcttaagaat tatcttaaaa datttcagaa aatttaattt 180
tactgttgtg ttttaggaaa aacgtattgc ataaagctat taatattgtc aggaaaagtg 240
cagagtagac taagaattag gaaaatteet agactaaaat nnnataagga gagggtttac 300
ctactgtttg aggcagttgg tctaatagta agcgattata gggagaaagc agaactactt 360
aactettetg tgttgaggaa tgacatgaaa ggtaggaaag gatataacaa atgetgataa 420
gaggagcctg atggatgaga ggagggaact gctttaaatg agttctactt cagacataag 480
ttaattctca gagccacaaa aactttcact ttcatttgtg aaatacaact cagttctcac 540
ggcttaacac tttaacccat ggagagacct gaagagttgg agaagcttgg cagatgcttc 600
                                                                   608
tgtgatag
<210> 125
<211> 402
```

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<212> DNA
<213> Banting cattle
<400> 125
gagagacatt atgacaccgc caaatttaat tgcagaggta agtataggta cacatattat 60
gttagataac ttgaagccaa cagtctaaat tttactgtca taccaataat gaataatctc 120
aagtattaag tgatatattt atottaaaga tggtotgaga aaatttgaaa ttaattttgo 180
tgttgtgttt ttggaaataa gtatcatgta aatgaggaag actaaattga attaactgaa 240
aactaggaga aatttataga ctaacagaat aaatagaggg ttatatctgt gatttgaggc 300
atttqqcatq ataqtaagaq attacaggga gaaaggagaa tggcttaatt ctgtaatgga 360
acatgacctg tacagtggaa aagggtataa tgaaatatgg at
<210> 126
<211> 479
<212> DNA
<213> Indian elephant
<221> misc feature
<222> (1)...(479)
\langle 223 \rangle n = A,T,C or G
<400> 126
gacattatga cnnnnnnnn nnnnnntgca gaggtaggta taaatgtttt atagtatgtt 60
qtataactta aaaccaaaag tctaaatatt actgccatag caatagtgaa tattctagat 120
tattaagtaa gataaatatt tatcttaagg atggtcttaa aaatttgagg gaaataaatt 180
taattttaat attatgtttt agaacaagta teecataace etatgagtaa tgtegtgaag 240
accaaaataa agaataggct aagaattagg agaaattcct aggataagaa taaaataagg 300
aaggggggca tgcctagtgt ttgaggcagt tggtgtaata ctaagagatt atatggagaa 360
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<210> 127
<211> 284
<212> DNA
<213> Fishing cat
<400> 127
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aacaaattta attttgctgt tgtgttttgg gaagcaagta tcctataaac ctgccggtac 240
taactagtag gaagactaat cccagagtag actaagaatt tgga
<210> 128
<211> 290
<212> DNA
<213> Sun bear
<220>
<221> misc feature
<222> (1)...(290)
<223> n = A, T, C or G
<400> 128
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ataatctcaa ttaattaagt ggaagtaaat tatttatctt aaagatggtc ttagacactt 180
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 <211> 272
 <212> DNA
 <213> Dwarf goat
 <400> 129
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 qttaqqtaac ttgaaqccaa cagtctaaat tttactgtca taccaataat gaataatcac 120
 aagtattaag taatattt atgttaaaga tggcctgaga aaatgtgaaa ttaactttgc 180
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 <211> 327
 <212> DNA
 <213> Guinea pig
 <220>
 <221> misc feature
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 <223> n = A, T, C or G
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  <212> DNA
  <213> Artificial Sequence
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  <223> Zoo43sUV
  <400> 131
                                                                     21
  tgtgctgaga gacattatga c
  <210> 132
  <211> 19
  <212> DNA
  <213> Artificial Sequence
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  <223> Zoo44aRV
  <400> 132
                                                                     19
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  <210> 133
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<212> DNA
<213> Man
<400> 133
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tgaattgctg caacatgatt gtcatcttca cttagccatt ggtcaagatc ttcacaaaag 180
ggtttgataa gttctagctg tggtgggtta tggtcttcaa aaggatattg cgcaactctg 240
taattagatt tggcggtgtc ataatgtctc tcagcacaac t
<210> 134
<211> 271
<212> DNA
<213> Chimpanzee
<400> 134
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tgcaacatga ttgtcatctt cacttagcca tcggtcaaga tcttcacaaa agggtttgat 180
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tttggcggtg tcataatgtc tctcagcaca a
<210> 135
<211> 271
<212> DNA
<213> Oran-utan
<220>
<221> misc_feature
<222> (1)...(271)
<223> n = A,T,C or G
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ctgcaacatg attgtcatct teacttagee attggtcaag atettcacaa aagggtttga 180
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<210> 136
<211> 268
<212> DNA
<213> Gorilla
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gctgcaacat gattgtcatc ttcacttagc cattggtcaa gatcttcaca aaagggtttg 180
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<210> 137
<211> 306
<212> DNA
<213> Domestic pig
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tctagctgtg gtggattatg atcttcaaaa ggatactgtg caactctgca gttaaatgtg 240
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<210> 138
<211> 258
<212> DNA
<213> Wild boar
<400> 138
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<213> SPL5 senseArtificial Sequence
<220>
<223> SPL5 sense
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<211> 20
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<223> Zoo44aRV antisense
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                                                                   20
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<211> 712
<212> DNA
<213> Man
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taaacccaaa atgaactgtc acatggacat ttcgtcagga ctctcaatgc aaaaggaata 600
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<210> 142
<211> 593
<212> DNA
<213> Chimpanzee
<220>
<221> misc feature
<222> (1)...(593)
<223> n = A, T, C or G
<400> 142
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<210> 143
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<213> Chimpanzee
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<221> misc feature
<222> (1)...(589)
\langle 223 \rangle n = A,T,C or G
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 tattgcttaa ctttctaatt gttaaataga aaatagtctc ttgataagtc ttaaatataa 420
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 <211> 593
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 tgattgtcat cttcacttag ccattggtca agatcttcac aaaagggttt gataagttct 180
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tgcatacaaa cttaacaggt atttaaaaaga aacactggat tccaaagaaa aataatgtat 360
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<212> DNA
<213> Orang-utan
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<400> 146
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 <211> 556
 <212> DNA
 <213> Domestic pig
 <400> 147
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<212> DNA
<213> Wild boar
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<212> DNA
<213> Cattle
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 <212> DNA
 <213> Banting cattle
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<212> DNA
<213> Sheep
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<210> 153
<211> 780
<212> DNA
<213> Bighorn
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<212> DNA
<213> Cameroon sheep
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agacaagtat tattttctca ctctwtgatg gaggcattgg agg
<210> 155
<211> 524
<212> DNA
<213> Deer
<400> 155
tetetggtee ttaetteece gtagaaatet agggeetett gtgeetttaa aaatttgeee 60
cgatgtaaca aatatgcaca aatcattaca ccagttcgtc cctttccagc tttacagtga 120
atcgctgcaa catgattgtc atcctcactt agccattggt caagatcttc acaaaagggc 180
ttgataagtt ctaactgtgg tggattatgg tcttcaaagg gatactgtgc aactgtgata 240
aaaaaatgac ctcagaataa gaaaataatt tcacttgaat tgcttattac aagtaggtta 300
actttagaaa tgttgcatat aaatagttta aaaatatccg aaccataaag aaaaagaatt 360
tattgtctgg taattttcta atttttgaac agaaaataat ctctcattaa ctcaaattta 420
tccattagaa aggtaagtca agtattgttt tcctcacttc atgatggagg caatggagga 480
gcaacatatc agaggcacag cataacagag gaagaggtaa actc
                                                                     524
<210> 156
<211> 647
<212> DNA
<213> Roe deer
<400> 156
tetetggtee ttaetteece gtagaaatet agggeetett gtgeetttaa aaatttgeee 60
cgatgtaaca aatatgcaca aatcattaca ccagttcgtc cctttccagc tttacagtga 120
atogotgoaa catgattgto atottoactt agocattggt caagatotto acaaaagggt 180
ttgataagtt ctaactgtgg tggattatgg tcttcaaagg gatactgtgc aactgtgata 240 aaaagataac ctcagaataa gaaaataatt tcacttgaat tgcttattac aagtaggtta 300
actttagaaa tgttgcatac aaatagttta aaaatatcca aaccataaag aaaagaattt 360
attgtctgat aattttctaa tttttgaaca gaaaataatc tcttatwaac tcaaatgtat 420
ccattagaaa ggtaagcaga gtattgtttt cctcacttca tgatgcaggc aatggaggag 480
caacatatca gaggtcacag cataacagag gaagaggtaa actcacaatg aaacatcaca 540
 tagectetta teaggaetet caatacattt tetageagaa ggaacegtaa tatetataac 600
```

```
attgcattat cacaaagtat gtattccaaa taaagtacat aacacta
                                                                  647
<210> 157
<211> 512
<212> DNA
<213> Goitred gazelle
<400> 157
teettaette eccatagaaa tetagggeet ettgtgeett taaaaatttg eccegatgta 60
acaaatatgc acaaatcatt acaccagttc gtccctttcc agctttacag tgaattgctg 120
caacatgatt gtcatcttca cttagccatt ggtcaagatc ttcacaaaag ggtttgataa 180
gttctaactg tggtggatta tggtcttcaa agggatactg tgcaactgtg ataaaaagat 240
aacctcagaa taagaaaata atctcacttg aattgcttat tataagtagg ttaactttat 300
aaatgttgca tacaaacagt ttaaaaaatat ctgaactaca gagaaaaaga atttattgtc 360
tgataatttc taattttttg acagaaaata atctctcata actcaaattt acccattaga 420
caggtaagcc aagtattatt ttctcacttt atgatggagg caatggagta gcacatatca 480
qaqqcacaac ctaacaqagg agaggtaact ca
<210> 158
<211> 798
<212> DNA
<213> Horse
<400> 158
ggtccttact tctccataga aatctagggc ctcctgtgcc tttaaaaact tgccccgatg 60
taacaaatat gcacaaatca ttacaccagt tcgtcccttt ccagctttac agtgaattgc 120
tgcaacatga ttgtcatctt cacttagcca ttggtcaaga tcttcacaaa agggtttgat 180
aagttctagc tgtggtggat tatgatcttc aaaaggatac tgtgcaactg tggtaaaaag 240
ataatctcaa attaagaaaa aaatctctcc tgaattgttt attaaaagta ggttaacttt 300
aggaatgctg cgtataagtt taacagatat ttaaaagaaa aactgaactc cagagaaaaa 360
taatttattg totgataatt ttotaatttt tgaatagaaa ataagagtoo cattaattot 420
caaaactcat ccattagaca gggaagccaa gtattatttt ccctactcta tgaaggagta 480
cattgtgcta tgcagaggta gcaaaggtca caacacataa gacatggagg tgaactcaaa 540
atgaaatgtc acatgggcct cttgttatgg ctttcaatgc atactctaac aaaaggagaa 600
ataacactta gaatattgca tcaccacaaa acatatattc caaagaaagt acagattact 660
aataaatcaa cggraaggat ggcattttac acttcatata ataaaaatgc taactgtgtt 720
attttaaaga tggtctggca aatggtagcg ctgtataccg actttaacag catttacaaa 780
qaaactqqaa aatcactt
<210> 159
<211> 519
<212> DNA
<213> African elephant
<221> misc feature
<222> (1)...(519)
<223> n = A, T, C or G
<400> 159
tggtccttac ttcnnnnnn nnnnnnnnn nnncttgtgc ctttaaaaat ttgccccgat 60
gtaacaaata tgcacaaatc attacaccag ttcgtccctt tccagcttta cagtgaattg 120
ctgcaacatg attgtcatct tcacttagcc attggtcaag atcttcacaa aagggtttga 180
taagctctag ttgtggtggg ttgtggtctt caaaaggata ctgtgcaact gtggtaaaaa 240
gataaactca gaataagaaa aaaatctctc ctgaattttt aattaaaagt aggttagctt 300
cagaaacatt gcacataaac tataaacagg tgtttaaata aaagataagc taaactccct 360
taaaaaaaaa tttattgcct gataacttgc tagtttttga atatagtctc tcactaactc 420
```

```
ttaaatgcat ccattaaaaa aggagaccaa gtattatttt ccccacatta tgctagagga 480
aactgtgtta tgctgaagta gcacaggtta catctcaga
<210> 160
<211> 776
<212> DNA
<213> Indian elephant
<220>
<221> misc feature
<222> (1)...(776)
\langle 223 \rangle n = A, T, C or G
<400> 160
tggtccttac ttccccataa aaatctaggg cttcttgtgc ctttaaaaaat ttgccccgat 60
gtaacaaata tgcacaaatc attacaccag ttcgtccctt tccagcttta cagtgaattg 120
ctgcaacatg attgtcatct tcacttagcc attggtcaag atcttcacaa aagggtttga 180
taagetetag ttgtggtggg ttgtggtett caaaaggata etgtgeaaet gtggtaaaaa 240
gataaactca gaataagaaa aaaatctctc ctgaattttt aattaaaagt aggttaqctt 300
cagaaacatt gcacataaac tataaacagg tgtttaaata aaagataagc taaactccat 360
taaaaaaaaa tttattgcct gataacttgc tagtttttga atatagtctc tcactaactc 420
ttaaatgcat ccattaaaaa aggagaccaa gtattatttt ccccacatta tgctagagga 480
aactqtqtta tqctqaaqta qcacaqqtta catctcagaq gtggagctga acccaaaaaq 540
aaatgttaca taggeetett gteaaggget gteaatgeat tttetaacaa aaggagtagt 600
gacactaata atattgcatc accttggtaa cagaacatat tctcaaaggt agaatggatt 660
attaacagaa tcagtaatgg aaaggattgc attttatact tcatataaaa natgttcggt 720
ctattattta aaggtggcct tacaaatgtt agtgttgtat acaatgattt ataaga
<210> 161
<211> 701
<212> DNA
<213> Dog
<400> 161
ggtccttact tccccataga aatctagggc ctcttgtgcc tttagaaatt tgccccgatg 60
taataaatat gcacaaatca ttacaccagt tcgtcccttt ccagctttac agtgaattgc 120
tgcaacatga ttgtcatctt cacttagcca ttggtcaaga tcttcacaaa agggtttgat 180
aagttctagc tgtggtggat tatggtcttc aaaaggatac tgtgcaactg tggtaaaaag 240
ataacctcag aattagaaaa aagtctttcc tgaactgttt attaaaagta ggttaacttt 300
agaaacattq catqtaaqct taacagatqt ttaaaagaaa aacggaactc cagagaaaaa 360
taatttgctg tctgataatt ttccaatttt tgaatagaaa atagtctctc attaattctt 420
aaacctacca ctagagagag aggctaagca ttattttccc cactttaatg aaagaggaaa 480
ctttgcaatg gagagggagc acacgtcaac atatcagagg gaagaggcaa actcaaaatg 540
aaatggcaca caggtttcct gtcagggctc tcaatgcatt ttctgacaaa aggagtcata 600
atatttataa tactacgtca tcacaaaata tatattccag agaaagtata aataaccgat 660
aaatcaatga tggaaaggat tgattttaca cttgatataa t
<210> 162
<211> 603
<212> DNA
<213> Sun bear
<220>
<221> misc feature
<222> (1)...(603)
<223> n = A, T, C \text{ or } G
```

```
<400> 162
ggtccttact tcnnnncata gaaatctagg gcctcttgtg cctttaaaaa tttgccccga 60
tgtaataaat atgcacaaat cattacacca gttcgtccct ttccagcttt acagtgaatt 120
gctgcaacat gattgtcatc ttcacttagc cattggtcaa gatcttcaca aaagggtttg 180
ataaqttcta gctqtqgtqg attatggtct tcaaaaggat actgtgcaac tgtggtaaaa 240
ggataacctc agaattagaa aaaagtcttt cctgaattgt ttattaaaga aggttaactt 300
taqaaatqtt qcatataaqc ttaacagatg tttaaaagaa aaactaaact ccagagaaaa 360
taatttgctg cctgacaatt tacgaatttt tgaatagaaa acagtctctc attaattctt 420
aaacccaccc acaagacaga ggccaagcat tatgttcccc acttaactga agaggaaaga 480
aactttgcta tggagaggta gcacaagtca catatcagag ggagaggcaa attcnaaatg 540
aaatgtcacg taggtaggtt tctgttaggg ctctcaatgc atttttctga caaaaggagt 600
cat
<210> 163
<211> 536
<212> DNA
<213> Mouse
<400> 163
ccttacttcc ccataaaaat ctagggcctc ttgtgccttt aaaaatttgc cccgatgcaa 60
taaatatgca caaatcatta caccagtccg tccctttcca gctttacagt gaattgctgc 120
aacatgattg tcatcttcac ttagccattg gtcaagatct tcacagaagg gtttgataag 180
ttctagctgt ggtgggttat ggtcttcaaa aggatactgt gcaactgttg caaaaagata 240
atcccagtgt aagaaaattt taaatttttt atttaaaaac ataggttaac tttcaaaatg 300
ttatatata acttactggt tcttaaaaga agcctaactt tcaggaaatt ttaatttatt 360
actaattaaa cotagatttt aaagaaagto tittattaat tottaaatgo attoattaga 420
catggaaaca agcattgtgc tcttcactcc agggaggatg aatctgtgca tgaagggaac 480
acgtcatage ctateagtee actgaateea aatgeaegte acceaggeae ttgtea
<210> 164
<211> 696
<212> DNA
<213> Guinea pig
<400> 164
acttetecat agaaatetag ageetettgt geetttaaaa atttgeeeeg atgtaataaa 60
tatgcacaaa tcattacacc agtccgtccc tttccagctt tacagtgaat tgctgcaaca 120
tgattgtcat cttcacttag ccattggtca agatcttcac aaaaaggctt gataagttct 180
agctgtggtg ggttatgatc ttcaaaaggg tattgtgcaa ctgtgataaa aacataatct 240
cagagtaaga aagggatett geetaaattt etaateagaa ataggteaae tttagaaatg 300
tttcacataa actcaagatg tttaaacaga aaaactgaac tgcatagaaa aataatttat 360
tgttcgttta cttttttact ttctttttt aaaatacaaa atagtctatt agtaactttt 420
aaacgtatct attacacaag gtggccaggt attacgttct tcacttcatg caggagaaaa 480
ctgtgatttg acagggaaca cagatcataa aacatcaaag atacatcgaa tccaaaaaaa 540
taccaggica cacagcetet cataacgict tiaggigaat tictgacaaa agcagtaaca 600
tttattatac tgcatcacca tacaacacac tttgaaggaa gtatgaacta ctaatrggat 660
acactatgaa aaarmtgcat tttatatttt ataaat
<210> 165
<211> 695
<212> DNA
<213> Himalaya-Tahr
<220>
<221> misc feature
<222> (1)...(695)
<223> n = A,T,C or G
```

```
<400> 165
acttennnnn nnnnnnnnn nnnnnnnnn nnnnnnnna atttgccccg atgtaacaaa 60
tatgcacaaa tcattacacc agttcgtccc tttccagctt tacagtgaat tgctgcaaca 120
tgattgtcat cttcacttag ccattggtca agatcttcac aaaagggttt gataagttct 180
aactgtggtg gattatggtc ttcaaaggga tactgtgcaa ctgtgataaa aagataaccg 240
caqaataaqa aaataatctc acttgaattg cttattacaa gtaggttaac tttagaaatg 300
ttqtatacaa ataqtttaaa aatatctgaa ctatagagga aaagaattta ttgtctgata 360
attttctaat tttgaacaga aaataatctc tcattaactc aaatttatcc attcgacagg 420
taagacaagt attetttee teactetatg atggaggeaa tggaggagea acatateaga 480
ggtcacaaca taacgsagga agaggcaaac tcaagagtga aacgtcgcac gagcctctta 540
traggertet craatargtt tertagraaa aggaartgta aratetataa tategratta 600
tcacaaaaca tgtattccaa agaaagtaca gatcactaat aggtccaatg cagaagactg 660
cattttatgt tgatgtgaca gaaaggcaaa gcata
<210> 166
<211> 281
<212> DNA
<213> Human
<400> 166
cettactice ceatagaaat etagggeete tigtgeetit aaaaattige eeegatgiaa 60
taaatatgca caaatcatta caccagttcg tccctttcca gctttacagt gaattgctgc 120
aacatgattg tcatcttcac ttagccattg gtcaagatct tcacaaaagg gtttgatcag 180
ttctagctgt ggtgggttat ggtcttcaaa aggatattgt gcaactgtgg taaaaagata 240
acctcagaat aagaaaaaaa actcctgaat ttttaattac a
<210> 167
<211> 373
<212> DNA
<213> Vikunja
<220>
<221> misc_feature
<222> (1) ... (373)
<223> n = A, T, C or G
<400> 167
caaatatgca caaatcatta caccagttcg tccctttcca gctttacagt gaattgctgc 120
aacatgattg tcatcttcac ttagccattg gtcaagatct tcacaaaaagg gtttgataag 180
ttctagctgt ggtggattat ggtcttcaaa aggatactgt gcaactgtgg ttaaaaaaaa 240
agaaaagaaa aaaagaacct cagaataaga aaaaaaatct cccctgaact gcttattaaa 300
tgcaagttaa ctttggaaat gttggcatat taaccttaac agacgtttta aaaggaaaat 360
                                                                 373
ctgaactcca gag
<210> 168
<211> 291
<212> DNA
<213> Spotted mustang
<220>
<221> misc feature
<222> (1)...(291)
<223> n = A, T, C or G
<400> 168
ctctggtcct tacttcccca tagaaatcta gggcctcttg tgcctttaaa aatttgcccc 60
```

```
gatgnaataa atatgcacaa atcattacac cagttcgtcc ctttccagct ttacagtgaa 120
ttgctgcaac atgattgtca tcttcactga gccattggtc aagatcttca caaaagggtt 180
tgataagttc cagctgcggt gggttatggt cttcaaaagg atactgtgca actgtgtaaa 240
aaqatcacct caqaqtqaqa aaaqaqtcct tcctgaactg tttcttaaaa g
<210> 169
<211> 598
<212> DNA
<213> Fishing cat
<400> 169
acttccccat agaaatctag ggcctcttgt gcctttaaaa atttgccccg atgcaataaa 60
tatgcacaaa tcattacacc agttcgtccc tttccagctt tacagtgaat tgctgcaaca 120
tgattgtcat cttcactgag ccattggtca agatcttcac aaaagggttt gataagttcc 180
agctgcggtg ggttatggtc ttcaaaagga tactgtgcaa ctgtgtaaaa agatcacctc 240
agaatgagaa aagaggcctt cctgaattgc ttcttaaaaag taggttaact tcagaaacgt 300
tgcatataag cttaacagat gtttagaagg aaaactaaac tccagagaaa aatactcgtc 360
tgatgatttt ccaatttttg aacagaaaac agtctctcat taatttttaa acctatgcac 420
tagacagaga ggccgattat ttccccccat gacgaagagg agactgctct ggagagcaag 480
cacaagtcac aacgtgtcag agggagagga ggacccggaa tgtcacacag gtttcctgtc 540
agggetetea atgeatttte tgacaaaatg agtaataege ttataetatt acateate
<210> 170
<211> 220
<212> DNA
<213> Turkey
<220>
<221> misc feature
<222> (1)...(220)
\langle 223 \rangle n = A,T,C or G
<400> 170
ctctggtcct tacttcccca tagaaatcta gggcttcttg agcctttaaa aatttgcctc 60
gatgtaataa atatgcacat atcattacac cagttcgtcc ctttccagct ttacagtgga 120
ttgctgcaac atgattgtca tcttcactta gccattggtc aagatcttca caaaanggtt 180
tgataagctc taactgtggt gggttatggt cttcaaaagg
<210> 171
<211> 505
<212> DNA
<213> Cockerel
<220>
<221> misc feature
<222> (1)...(505)
<223> n = A, T, C or G
<400> 171
totggtcctt acttocccat agaaatotag ggcttcttga gcctttaaaa acttgcctcg 60
atgcaacaaa tatgcacata tcattacacc agttcgtccc tttccagctt tacagtggat 120
tgctgcaaca tgattgtcat cttcacttag ccattggtca agatcttcac aaaaaggttt 180
gataagetet aactgiggtg ggttatggte tteaaagggg taetgtgeaa etgtaaigag 240
aaggattaac ttattaaaat ctaaaaagga taatcaccaa gagctcaact agacaggtca 300
aatttgtgac aagcatgaaa aaattaacat totaaataca gtottgcata tagatttgta 360
tacacgcaac tttgattctg ctgttattca gtaacattgc acactaaatg catcacaaat 420
ttctctagta atacgtaagt atcttactgg catgaagagg actatcccac cgtgcttctg 480
```

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505
nagttnntac tacagactct gcacc
<210> 172
<211> 645
<212> DNA
<213> Duck
<220>
<221> misc_feature
<222> (1) ... (645)
<223> n = A, T, C or G
<400> 172
cettacttee ceatagaaat etagageette ttgageettt aaaaaettge etetatgeaa 60
cagatatgcg catatcatta caccagttcg tccctttcca gctttacagt ggattgctgc 120
aacatgattg tcatcttcac ttagccattg gtcaagatct tcacaaaaag gtttaatgag 180
ctcaagctgt ggtgggttat ggtcttcaaa agggtactgt gcaactgcaa caagaaagaa 240
aaacttacca aaatctcaaa aggaaactac agcaagcttg actagacgtg tcatctttgg 300
acaagcacac acaaaaatta acattctaaa taaaaactgt cttatatgta tatacatata 360
gctttgcttt cactgttatt cagcagcata ctatacactn ttncacatca cagacatttc 420
totattaata cataagcaca tatottagac tatttoacag tgottotgaa acaagtogca 480
cagactetat tttacactat ttttctgaaa tttaagagtg cactggcaca aagaataacc 540
ttgtgaaaac ccattagtca cagactacct gctgagagaa agcagggcaa acctcactca 600
ctgatcagag acagggattt tgcagcaaga attctgagtg gctgg
                                                                   645
<210> 173
<211> 516
<212> DNA
<213> Quail
<220>
<221> misc feature
<222> (1)...(516)
<223> n = A, T, C or G
<400> 173
ccttacttcn nnnnnnnnn nnnnnnnnn nnnnnccttt aaaaacttgc ntcgatgcaa 60
caaatatgca catatcatta caccagttcg tccctttcca gctttacaat ggattgctgc 120
aacatgattg tcatcttcac ttagccattg gtcaagatct tcacaaaaag gtttgataag 180
ctctagctgt ggtgggttat ggtcttcaaa agggtactgt gcaactgcaa tgagaaggaa 240
taacqttcta aataaaacac agtettgcat acagatttgc atccacacag ctttgattct 300
gttgttattc agcagcatat tacacactat aaatgcatca catgtttctc tagtaatacg 360
taagcatctt gctgcatgaa gagacctcag aagcattgtg ggaatagtta gtgctaccaa 420
ctgcacctta caccatgatt ttactcaaat taagagtgta ctggcacaaa aaataacgtg 480
                                                                    516
ttttaaggtc acccatcaaa tgcagtgtct tttttt
<210> 174
<211> 395
<212> DNA
<213> Trout
<220>
<221> misc feature
<222> (1)...(395)
<223> n = A, T, C or G
<400> 174
```

```
tctctqqtcc ttacttcnnn nnnnnnnnn nnnnnnnnn ngctttgagg aacttgcccc 60
ggtgtaacag gtaagcacag atcatgacac ccgtacgtcc ctttccagct ttacagtgaa 120
tegeegeeac atgattgteg tetteaetta accaaaggte aagatetteg cagaaeggtt 180
tgatcagete cagetgggge ggattgtgat ceteaaaegg atattgtgea actggagana 240
gacagacaga gaccgggctc agttagttag cgtcacacgt gggtttttag tgaaagattg 300
atteatteae tgactgeetg aaagacagtg ataatggttt cactetgatg taatatetaa 360
cctctqcaat tqaatttqtq ttqcqtcata atqtc
<210> 175
<211> 21
<212> DNA
<213> Artificial Sequence
<220>
<223> PTENse sense
<400> 175
                                                                   21
atcttgacca atggctaagt g
<210> 176
<211> 20
<212> DNA
<213> Artificial Sequence
<220>
<223> Zoo44aRV
<400> 176
                                                                   20
ttgtctctgg tccttacttc
<210> 177
<211> 160.
<212> DNA
<213> Goat
<400> 177
tetetggtee ttaetteece atagaaatet agggeetett gtgeetttaa aaatttgeee 60
cqatqtaaca aatatqcaca aatcattaca ccagttcqtc cctttccagc tttacagtga 120
attgctgcaa catgattgtc atcttcactt agccattggt
<210> 178
<211> 150
<212> DNA
<213> Antelope
<220>
<221> misc feature
<222> (1)...(150)
<223> n = A, T, C or G
<400> 178
ctqqtcctta cttccccata qaaatctagg gcctnntgtg cctttaaaaa tttgccccga 60
tgtaacaaat atgcacaaat cattacacca gttcgtccct ttccagcttt acagtgaatt 120
gctgcaacat gattgtcatc ttcacttagc
<210> 179
<211> 153
```

<212> DNA <213> Kanga	roo			
cgatgtaata	aatatgcaca	ccagttcgtc	aaactttcct tttacagtga	
<210> 180 <211> 154 <212> DNA <213> Rabbi	.t			
ccgatgtaat	aaatatgcac	accagttcgt	aaaatttgcc ctttacagtg	
<210> 181 <211> 155 <212> DNA <213> Hare				
taataaatat	gcacaaatca	tcgtcccttt	tgccccgatg agtgaattgc	
<210> 182 <211> 159 <212> DNA <213> Goose	e)			
cgatgcaaca	aatatgcgca	ccagttcgtc	aaacttgcct tttacagtgg	60 120 159
<210> 183 <211> 156 <212> DNA <213> Ostri	ich			
gatgťaacaa	ataagcacat	cagttcgtcc	aacttgcctc ttacagtgga	
<210> 184 <211> 151 <212> DNA <213> Pigeo	on			
atgcaacaaa	tatgcacata	agttcgtccc	acttgcctcg tacagtggat	

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<210> 185
<211> 163
<212> DNA
<213> Varan
<400> 185
tctctggtcc ttacttcccc atagaaatct agagcttctt gtgccttttg aaatcttcct 60
cgatgtaata aatatgcaca aatcattaca ccagttcgtc cctttccagc tttacaatgg 120
attgccgcaa cgtgattgcc atcttcactt agccattggt caa
<210> 186
<211> 160
<212> DNA
<213> Trout
<400> 186
{\tt tctggtcctt}\ acttcaccgt\ agaagtccag\ agcttcctgt\ gctttgagga\ acttgcccg\ 60
gtgtaacagg taagcacaga tcatgacacc cgtacgtccc tttccagctt tacagtgaat 120
cgccgccacg tgattgtcgt cctcacttag ccattggtca
<210> 187
<211> 23
<212> DNA
<213> Artificial Sequence
<220>
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<210> 188
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<223> PTENex6R antisense
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gcaagttccg ccactgaa
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catctggatt atagaccagt ggcactgttg tttcacaaga tgatgtttga aactattcca 120
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atgttcagtg gcggaact
<210> 190
<211> 131
<212> DNA
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<210> 196				

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gtggcggaac t
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<212> DNA
<213> Goitred gazelle
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<211> 132
<212> DNA
<213> Horse
<400> 198
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gattatagac cagtggcact gttgtttcac aagatgatgt ttgaaactat tccaatgttc 120
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agtggcggaa ct
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<213> Dog
<400> 199
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cggaa
<210> 200
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<212> DNA
<213> Sun bear
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gtggcggaa
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<212> DNA
<213> Rabbit
<400> 201
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	g aggegetatg g gecetgetgt					60 120 127
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	a gtcagaggcg c ccgtggcatt a ct					
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gattacagac agtggcggaa	cagtggcact ct	gctgtttcac	aagatgatgt	ttgaaacaat	teccatgtte	120 132
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ggcggaactt
<210> 215
<211> 122
<212> DNA
<213> Carp
<400> 215
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ct
<210> 216
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<212> DNA
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tggcggaact
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<211> 132
<212> DNA
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agtggcggaa ct
<210> 218
<211> 129
<212> DNA
<213> Tench
<400> 218
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1 3

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<212> DNA
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<211> 25
<212> DNA
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<220>
<223> PTENex7R antisense
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<211> 168
<212> DNA
<213> Man
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<221> misc_feature
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atatcaaagt agagttette cacaaacaga acaagatget aaaaaagg
<210> 222
<211> 159
<212> DNA
<213> Chimpanzee
<400> 222
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gggaagacaa gttcatgtac tttgagttcc ctcagccgtt acctgtgtgt ggtgatatca 120
aagtagagtt cttccacaaa cagaacaaga tgctaaaaa
<210> 223
<211> 161
<212> DNA
<213> Cattle
<400> 223
cagtttgtgg tctgccagct aaaggtgaag atatattcct ccaattcagg acccacacga 60
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<213> Sheep
<400> 224
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ggaagacaag ticatgtact tigagttccc tcagccgctg cctgtgtgtg gtgacatcaa 120
agtagagttc ttccacaaac agaacaagat gctaaaaaag
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<210> 225
<211> 161
<212> DNA
<213> Goat
<400> 225
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aaagtagagt tcttccacaa acagaacaag atgctaaaaa a
<210> 226
<211> 153
<212> DNA
<213> Red buffalo
<400> 226
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aagtagagtt cttccacaaa cagaacaaga tgc
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<210> 227
<211> 159
<212> DNA
<213> Deer
<400> 227
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cgggaagaca agttcatgta ctttgagttc cctcagccgt tgcctgtgtg tggtgacatc 120
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aaagtagagt tcttccacaa acagaacaag atgctaaaa
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<211> 162
<212> DNA
<213> Roe deer
<400> 228
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<210> 229
<211> 161
<212> DNA
<213> Goitred gazelle
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                                                                    160
caaagtagag ttcttccaca aacagaacaa gatgctaaaa
<210> 236
<211> 163
<212> DNA
<213> Kangaroo
<400> 236
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ggcgggaaga caagcacatg tacttcgagt tcccccagcc tctgccggtg tgtggcgaca 120
ttaaagtgga attcttccac aaacagaaca agatgctaaa aaa
<210> 237
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\langle 223 \rangle n = A,T,C or G
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aaagnggagt tcttccacaa acaga
<210> 238
<211> 146
<212> DNA
<213> Chicken
<400> 238
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cgtgaagaca agtatatgta ctttgaattc cctcaacctt tgccggtatg cggtgatatc 120
aaagtggagt tcttccacaa acagaa
<210> 239
<211> 154
<212> DNA
<213> Duck
<400> 239
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cgtgaagaca agtatatgta ctttgaattc cctcaacctt tgccggtatg cggtgatatc 120
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<213> Quail
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acgtgaagac aagtatatgt actttgaatt ccctcaacct ttgccggtat gcggtgatat 120
caaagtggag ttcttccaca aacagaacaa gatgctaaaa aag
<210> 241
<211> 160
<212> DNA
<213> Ostrich
<400> 241
gtttgtggtc tgccagctaa aggtaaagat attcacctcc ccttcaggac cctcaagacg 60
tgaagacaag tatatgtact ttgaattccc tcaacccttg ccggtatgcg gtgatatcaa 120
agtggaattc ttccacaaac agaacaagat gctaaaaaag
<210> 242
<211> 145
<212> DNA
<213> Pigeon
<400> 242
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acgtgaagac aagtatatgt actttgaatt ccctcaacct ttgccggtat gcggtgatat 120
caaagtggaa tttttccaca aacag
<210> 243
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gcgagaggag aagtacatgt acttngattt tccncagenn ctgcctgtgt gnggagacat 120
caaggtggag ttcttccaca aacagaacaa gatgctaaaa aag
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<220>
<221> misc feature
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<223> n = A,T,C or G
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<400> 250
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<210> 251
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<212> DNA
<213> Sheep
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<212> DNA
<213> Goat
<400> 252
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gataatgaca aggaatatet agtaeteaet ttaacaaaaa atgatettga caaagcaaat 180
                                                                   213
aaagacaagg ccaaccgata cttttctcca aat
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<211> 212
<212> DNA
<213> Red buffalo
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aaagacaagg ccaaccgata cttttctcca aa
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<211> 213
<212> DNA
<213> Deer
<400> 254
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ataatgacaa agaatatcta qtactcactt taacaaaaaa tgatctcgac aaagcaaata 180
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<211> 214
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<213> Roe deer
<400> 255
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gataatgaca aagaatatot agtactcact ttaacaaaaa atgatotoga caaagcaaat 180
aaagacaagg ccaaccgata cttttctcca aatt
<210> 256
<211> 213
<212> DNA
<213> Goitred gazelle
<400> 256
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gataatgaca aggaatatet agtacteaet ttaacaaaaa atgatetega caaagcaaat 180
aaagacaagg ccaaccgata cttttctcca aat
<210> 257
<211> 213
<212> DNA
<213> Horse
<400> 257
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gtagaaaatg gaagtctatg tgatcaagaa attgatagta tttgcagtat agagcgtgca 120
gataatgaca aagaatatet agtacteaet ttaacaaaaa atgatetega caaageaaat 180
aaagacaagg ccaaccgata cttttctcca aat
<210> 258
<211> 210
<212> DNA
<213> Dog
<400> 258
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aatgacaagg aatatctagt actcacttta acaaaaaatg atctcgacaa agcaaataaa 180
gacaaggcca accgatactt ttctccaaat
<210> 259
<211> 213
<212> DNA
<213> Sun bear
<400> 259
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<211> 210
<212> DNA
<213> Rabbit
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<210> 261 <211> 210 <212> DNA <213> Hare						
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<210> 262 <211> 203 <212> DNA <213> Antel	Lope					
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<210> 263 <211> 213 <212> DNA <213> Kanga	aroo					
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caaagccaac cgatactttt ctccaaat
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<211> 213
<212> DNA
<213> Chicken
<400> 266
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aatgacaagg aatatttaat ccttacatta acaaaaaatg atctagacaa agcaaataaa 180
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acaaagccaa ccgatacttt tctccaaatt
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<213> Goose
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gataatgata aggaatattt aateettaca ttaacaaaaa atgatetaga caaagcaaat 180
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<213> Dog
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<212> DNA
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<213> Rabbit
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